

Synthesis of Biological Markers in Fossil Fuels. 7.¹ Selected Diastereomers of 4 α -Methyl-5 α -stigmastane and 5 α -Dinosterane

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Received December 23, 1992

Efficient routes for the preparation of selected C-23 and C-24 diastereomers of the C₃₀ biological markers 4 α -methyl-5 α -stigmastane (1) and 5 α -dinosterane (2) involved the alkylation of 20-(iodomethyl)-4 α -methyl-5 α -pregnane with either saturated or α,β -unsaturated esters. The alkylation of (20S)-20-(iodomethyl)-4 α -methyl-5 α -pregnane with methyl (3R)-3-ethyl-4-methylpentanoate furnished methyl (20R,23 ζ ,24S)-4 α -methyl-5 α -stigmastane-23-carboxylate, and a subsequent decarbomethoxylation provided (20R,24R)-1. The alkylation of (20S)-20-(iodomethyl)-4 α -methyl-5 α -pregnane with methyl (3S)-3,4-dimethylpentanoate led to methyl (20R,23 ζ ,24R)-4 α ,24-dimethyl-5 α -cholestane-23-carboxylate, and the reduction of this mixture provided principally (20R,23S,24R)-5 α -dinosteran-29-ol. The further reduction of the mesylate of this isomer secured (20R,23S,24R)-5 α -dinosterane (2a). The application of the same sequence of reactions using methyl (3R)-3,4-dimethylpentanoate led principally to (20R,23R,24S)-5 α -dinosterane (2d). The alkylation of (20S)-20-(iodomethyl)-4 α -methyl-5 α -pregnane with methyl (2 ζ)-3,4-dimethyl-2-pentanoate and a subsequent reduction of the ester provided a separable mixture of (20R,23R)- and (20R,23S)-5 α -dinoster-24-(28)-en-29-ol in a 2.4:1 ratio. The conversion of (20R,23R)-5 α -dinoster-24-(28)-en-29-ol to the corresponding *tert*-butyldimethylsilyl ether, reduction of the $\Delta^{24(28)}$ bond with hydrogen over platinum oxide, and deprotection gave principally (20R,23R,24R)-5 α -dinosteran-29-ol. The further reduction of this alcohol provided (20R,23R,24R)-5 α -dinosterane (2b). The application of the same sequence of reactions to (20R,23S)-5 α -dinoster-24-(28)-en-29-ol provided (20R,23S,24S)-5 α -dinosterane (2c). Diastereoselectivity at the C-23 position in these ester alkylations was examined as a function of stereochemistry at both the C-20 and C-24 positions.

The formation of petroleum from biological materials under the conditions of high temperature and pressure over the course of geologic time results in the conversion of functionalized, terpenoid natural products to their corresponding hydrocarbon skeletons.² These "molecular fossils", or biological markers,² provide considerable information on the origin, maturation, migration, and microbiological degradation of crude oils. Among the many saturated steranes in crude oils in the C₂₇ to C₃₀ range, the C₃₀ 4 α -methyl-5 α -steranes such as 4 α -methyl-5 α -stigmastane³ (1) and 5 α -dinosterane⁴ (2) presumably derive from 4 α -methylsterols in marine algae and represent characteristic biomarkers diagnostic of marine origins. Among the present day representatives of these skeletons are the C₃₀ biosynthetic intermediates that are involved in the formation of plant sterols such as β -sitosterol (3). In addition, there are C₃₀ sterols⁵ such as dinosterol⁶ (4) and related analogs that are isolated from unicellular algae

in the phylum, *Pyrrhophyta*. In order to confirm the stereochemistry of specific members of 4 α -methyl-5 α -stigmastane³ (1) and 5 α -dinosterane^{4,7} (2) families detected by GC-MS techniques in crude oils,² we devised a synthesis of selected diastereomers of these C₃₀ biomarkers.

As shown in Scheme I, a retroanalysis of the biomarker targets 1 and 2 suggested a 24-alkyl 4 α -methyl-5 α -cholestane-23-carboxylate (5) as a common precursor. Further disconnection of the ester 5 suggested either the alkylation of a lithocholate ester⁸ 6 with an acyclic electrophile 7 (path a) or the alkylation of an acyclic ester 9 with a suitable electrophilic sterane 8 (path b) as logical disconnections of 5. Of these two options, the latter path offered the best opportunity to control stereochemistry at the C-20, C-23, and C-24 positions, and the alkylation of

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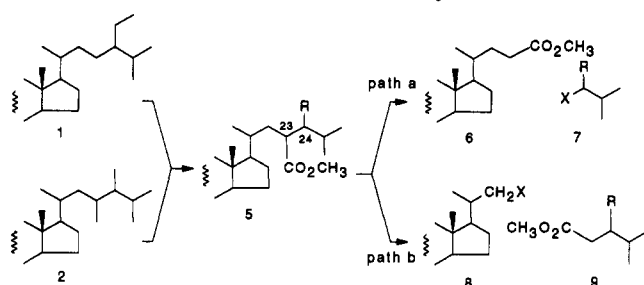
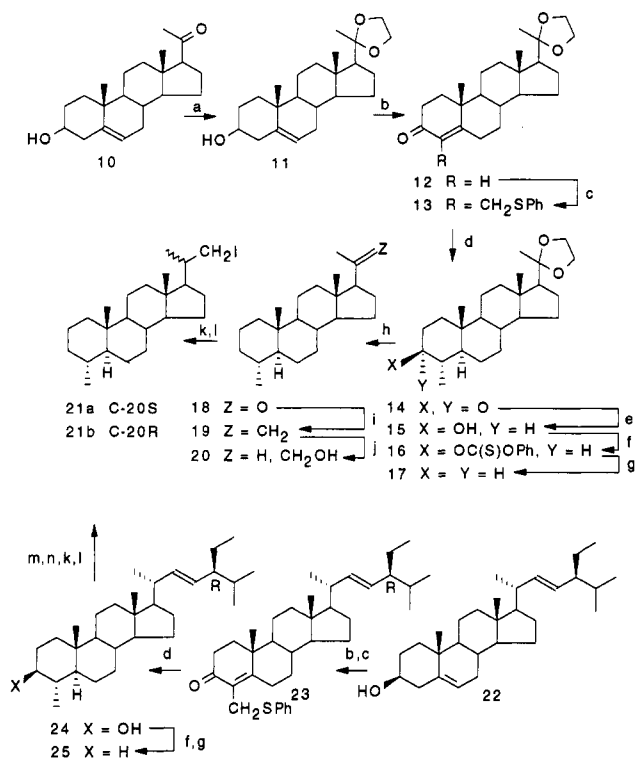
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Scheme I. Retroanalysis

Scheme II. Synthesis of 20-(Iodomethyl)-4 α -methyl-5 α -pregnane (21a)^a

^a Key: (a) HOCH₂CH₂OH, *p*-TsOH; (b) Al(O-*i*-Pr)₃, 1-methyl-4-piperidone; (c) PhSH, HCHO, Et₃N; (d) Li, NH₃; (e) LiAlH₄, THF; (f) PhOC(S)Cl, Py; (g) (*n*-Bu)₃SnH, AIBN, C₆H₆; (h) 1:2.5:5.5 HCl-HOAc-THF, (i) Ph₃P=CH₂, KO-*t*-Am, benzene; (j) BH₃·THF; NaOH, H₂O₂; (k) Et₃N, MsCl; (l) NaI, acetone; (m) O₃, Me₂S followed by NaOH, MeOH; (n) NaBH₄.

either saturated⁹ or α,β -unsaturated acyclic esters¹⁰ with sterane electrophiles successfully provided stereoselective routes to the biomarkers 1 and 2.

According to the retroanalysis, the synthesis of (20*R*, 24*R*)-4 α -methyl-5 α -stigmastane (1) required the electrophilic sterane, (20*S*)-20-(iodomethyl)-4 α -methyl-5 α -pregnane (21a), and methyl (3*R*)-3-ethyl-4-methylpentanoate.¹¹ As shown in Scheme II, the preparation of (20*S*)-20-

(8) The alkylation of methyl (20*R*)-5 β -cholanate or the alkylation of methyl (20*R*)-23-carbomethoxy-5 β -cholanate with a suitable alkyl bromide did not provide an efficient pathway for side-chain construction. The enolate Claisen rearrangement of (2'*E*)-2'-methyl-2'-butenyl 5 β -cholanate also failed to produce acceptable yields of the desired C-23 alkylated products. Finally, when efforts to introduce the C-4 α methyl and invert the C-5 β stereochemistry in cholanate systems proceeded in low overall yields, path a in Scheme I was abandoned.

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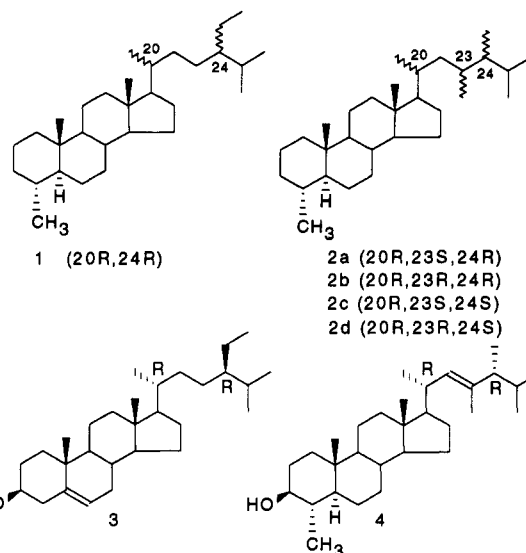


Figure 1.

(iodomethyl)-4 α -methyl-5 α -pregnane (21a) from pregnenolone (10) required the introduction of the 4 α -methyl group and a one-carbon homologation of the C-20 ketone. The ketalization of pregnenolone¹² (10), Oppenauer oxidation¹³ of the homoallylic alcohol 11, and a Kirk-Petrow thiophenoxymethylation¹⁴ of the enone 12 provided the α -(thiophenoxymethyl) enone 13. The lithium in ammonia reduction of 13 effected both desulfurization and enone reduction to furnish the ketal 14 having the 5 α -stereochemistry and the 4 α -methyl group.¹⁵ Deoxygenation¹⁶ at C-3 involved the further reduction of 14 to the alcohol 15, conversion to the phenyl thiocarbonate 16, and reduction of 16 with tri-*n*-butyltin hydride in order to secure the ketal 17. The acid-catalyzed hydrolysis¹⁷ of 17 to the ketone 18, a Wittig reaction, and a hydroboration-oxidation of the olefin 19 delivered the alcohols 20 as a separable C-20 epimeric mixture. The individual conversion of these epimeric alcohols to the corresponding mesylates¹⁸ and S_N2 substitution with sodium iodide¹⁹ provided (20*S*)- and (20*R*)-20-(iodomethyl)-4 α -methyl-5 α -pregnane (21a) and (21b). As shown in Scheme II, the application of a similar sequence of reactions to (22*E*)-stigmastanol (22) provided an alternate source of 21a and 21b. Although both 21a and 21b were used in alkylation studies that will be described later in the paper, only the

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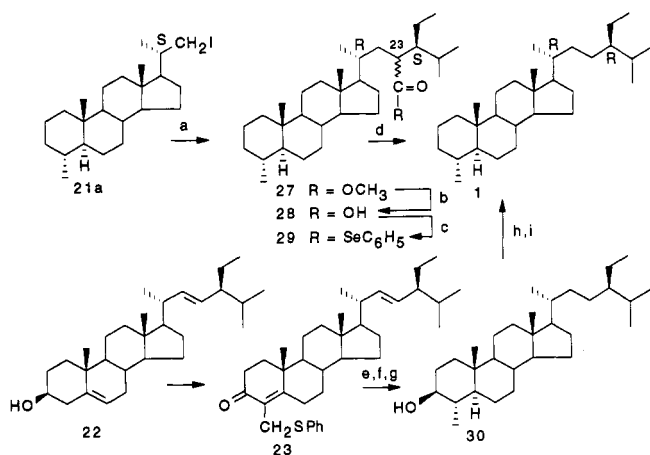
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(17) Vigorous hydrolysis conditions (i.e., refluxing for 17 h in 1:2:3.5 1 M HCl-HOAc-THF) resulted in epimerization at C-17 to give a 4:1 ratio of 18 and 4 α -methyl-5 α ,17 β (H)-pregnan-20-one: mp 124–126.5 °C; IR (CHCl₃) 1690 (C=O) cm⁻¹; ¹H NMR (CDCl₃) δ 0.78 (s, 3, C-18 CH₃), 0.79 (d, *J* = 6 Hz, 3, C-4 α CH₃), 0.90 (s, 3, C-18 CH₃), 2.12 (s, 3, C-21 CH₃); ¹³C NMR (CDCl₃) δ 12.9, 20.3, 20.7, 21.6, 24.0, 25.6, 31.0, 32.2, 32.6, 35.1, 35.3, 36.2, 38.6, 45.5, 50.3, 53.1, 53.8, 61.2, 213.2. Anal. Calcd for C₂₂H₃₈O: C, 83.48; H, 11.47. Found: C, 83.35; H, 11.46.

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**Scheme III. Synthesis of
(20*R*,24*R*)-4 α -Methyl-5 α -stigmastane (1)^a**



^a Key: (a) LDA, methyl (3*R*)-3-ethyl-4-methylpentanoate (26), THF, HMPA, -78 °C; (b) LiI, collidine, heat; (c) PhOP(O)Cl₂, Et₃N, PhSeH, Py; (d) (*n*-Bu)₃SnH; (e) Li, NH₃; (f) LiAlH₄, THF; (g) H₂, 10% Pd-C; (h) PhOC(S)Cl, Py, CH₂Cl₂; (i) (*n*-Bu)₃SnH, AIBN, C₆H₆.

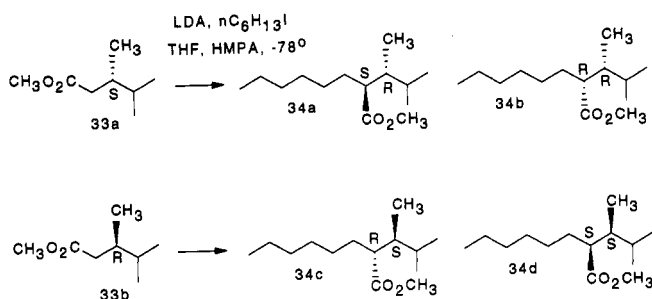


Figure 2.

(20*S*)-isomer 21a was used in the synthesis of specific biomarkers.

As shown in Scheme III, the alkylation of methyl (3*R*)-3-ethyl-4-methylpentanoate¹¹ (26) with (20*S*)-20-(iodomethyl)-4 α -methyl-5 α -pregnane (21a) furnished methyl (20*R*,23 ζ ,24*S*)-4 α -methyl-5 α -stigmastane-23-carboxylate (27a) and (27b) as a mixture of C-23 epimers. Optimal conditions for the alkylation involved the use of a 7.5:5:5:1 ratio²⁰ of diisopropylamine, *n*-butyllithium, ester 26, and iodide 21a at temperatures of ca. -30 °C. It was interesting that this approach for constructing the C22,23 bond was more satisfactory than other alternatives including the condensation of a Grignard reagent derived from 1-bromo-2-ethyl-3-methylbutane with (20 ζ)-4 α -methyl-5 α -pregnane-20-carboxaldehyde or the condensation of a Grignard reagent derived from (20 ζ)-20-(iodomethyl)-4 α -methyl-5 α -pregnane (21) with 2-ethyl-3-methylbutanal.

Lithium iodide in refluxing collidine²¹ effected saponification of the hindered carbomethoxy group in 27. The conversion of the intermediate carboxylic acid 28 to the phenylseleno ester²² 29 and reduction with tri-*n*-butyltin hydride provided (20*R*,24*R*)-4 α -methyl-5 α -stigmastane (1). An authentic sample of (20*R*,24*R*)-4 α -methyl-5 α -

stigmastane (1) was also prepared from (22*E*)-stigmasterol (22) using the route summarized in Scheme III. The identity of 1 prepared from either 21a or from 22 confirmed the C-20 stereochemical assignment of 21a and set the stage for the preparation of selected C-20*R* diastereomers of 5 α -dinosterane (2).

Previous efforts²³ to prepare 5 α -dinosteranes (2) from dinosterol (4) were hampered by the limited supplies of 4, the C-24 allylic epimerization during the hydrogenation of 4, and the difficulties associated with the reversed-phase HPLC separation of the C-23,24 diastereomers of 2. In order to prepare appreciable quantities of selected 5 α -dinosteranes (2) by the route suggested in Scheme I, we needed to define the factors that controlled C-23 diastereoselection in the alkylation of chiral esters with chiral sterane electrophiles. To this end, we completed a series of studies involving the alkylation of chiral esters with achiral electrophiles, the alkylation of achiral esters with (20*S*)- and (20*R*)-20-(iodomethyl)-4 α -methyl-5 α -pregnane (21a) and (21b), and finally the alkylation of chiral esters with 21a and 21b.

We began by investigating the diastereoselectivity of alkylations of chiral esters with achiral electrophiles. An enantioselective reduction of methyl (2*E*)- or (2*Z*)-3,4-dimethyl-2-pentenoate (32a) or (32b) with sodium borohydride in conjunction with a cobalt-semicorrin complex²⁴ provided methyl (3*S*)- or (3*R*)-3,4-dimethylpentanoate (33a) and (33b), respectively. As shown in Figure 2, the alkylation of ester 33a with an achiral iodide, *n*-hexyl iodide, led to a 4:1 mixture of the erythro- and threo-diastereomers 34a and 34b, respectively. In a similar fashion, the alkylation of ester 33b led to a 4:1 mixture of the erythro- and threo-diastereomers 34c and 34d. The optimal conditions for this alkylation again involved the use of a 7.5:5:5:1 ratio of diisopropylamine, *n*-butyllithium, ester 33a or 33b, and *n*-hexyl iodide at temperatures of ca. -30 °C. An alkylation performed with an equimolar ratio of these reagents led to a similar ratio of products and excluded the possibility that the excess of the ester enolate present in the former alkylation promoted equilibration of the erythro- and threo-products.

The erythro-selectivity was consistent with a model involving S_N2 attack by the *Z*-enolate of either 33a or 33b on *n*-hexyl iodide. As shown in Figure 3, this model assumed that an unfavorable, intramolecular steric interaction between the C-3 methyl group and the lithium alkoxide of the enolate disfavored conformers C and D in which the incoming electrophile approached from the side opposite the bulky isopropyl group. The absence of this unfavorable steric interaction in conformers A and B favored the erythro products 34a and 34c. This model differed from the Felkin-Anh model in which the principal steric interaction involved the incoming nucleophile and the proximal substituent on the adjoining chiral center. That is, a strict application of the Felkin-Anh model to the conformers in Figure 3 would predict that conformers C and D would be favored over conformers A and B since

(23) PCC oxidation of a sample of dinosterol (4), contaminated with (24*R*)-4 α ,24-dimethyl-5 α -cholestan-3 β -ol, followed by a Wolff-Kishner reduction furnished (22*E*)-5 α -dinoster-22-ene. Catalytic hydrogenation using palladium on carbon proceeded with epimerization^{6e} at C-24 and provided (20*R*,23 ζ ,24 ζ)-5 α -dinosterane (2) as a mixture of four diastereomers that were separated by reversed-phase HPLC. No epimerization at the other allylic C-20 position was observed during these catalytic hydrogenations.^{6e}

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(20) 20-(Iodomethyl)-4 α -methyl-5 α -pregnane is sparingly soluble at -78 °C in 6% HMPA-THF mixtures. An alkylation performed and quenched at -78 °C led principally to unreacted iodide and products derived from a Claisen condensation of the ester.

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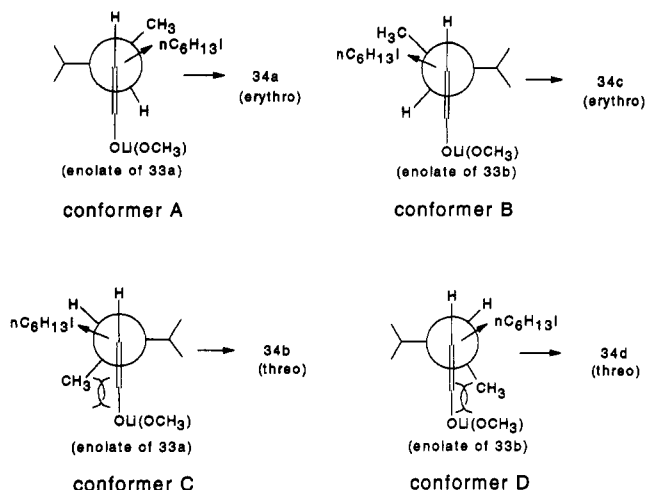


Figure 3.

the former minimized steric interactions between the *n*-hexyl iodide and the proximal C-3 methyl group. The model proposed here discounted this steric interaction as less important than the interaction of the lithium alkoxide and the C-3 methyl group.

The influence of the chirality of the C-20 stereocenter in the sterane electrophiles 21a and 21b on C-23 diastereoselectivity was examined in alkylations with achiral, saturated and unsaturated esters: methyl pentanoate (35), methyl 4-methylpentanoate (36), and methyl (2 β)-3,4-methyl-2-pentenoate²⁵ (32). The alkylation of 21a with the saturated ester, methyl pentanoate (35) (entry 1, Table I), exhibited a 5.1:1 preference for the " α -oriented" epimer 37a relative to the " β -oriented" epimer 37b. The alkylation of methyl pentanoate (35) with the enantiomeric electrophile 21b (entry 2, Table I) exhibited a similar 5.3:1 preference for the " α -oriented" epimer 37c relative to the " β -oriented" epimer 37d. In the same fashion, the alkylation of 21a with methyl 4-methylpentanoate (36) (entry 3, Table I), exhibited a 6.6:1 preference for the " α -oriented" epimer 38a relative to the " β -oriented" epimer 38b. Finally, the alkylation of 21a or 21b with the α,β -unsaturated ester, methyl 3,4-dimethyl-2-pentenoate (32) (entries 4 and 5, Table I), also displayed an approximate 2:1 preference for the " α -oriented" epimers 39a or 39c relative to the " β -oriented" epimers 39b or 39d, respectively. In summary, the chirality of the sterane electrophile favored " α -selectivity" at C-23 to a slight degree, but changing a single chiral center at C-20 in the electrophile, despite its proximity to the electrophilic terminus, had little influence. This finding was consistent with other alkylations²⁶ in which distal, acyclic, chiral centers had limited impact on diastereoselectivity.

Finally, it was of interest to examine the outcome of alkylations in which the "erythro-selectivity" that was noted in the alkylation of achiral electrophiles with chiral esters (Figure 2) was juxtaposed with the " α -selectivity" noted in the alkylation of chiral electrophiles 21a and 21b with achiral esters (entries 1–5, Table I). As shown in Table I, the alkylation of methyl (3*R*)-3,4-methylpentanoate (33b) (entries 6 and 7, Table I) with 21a or 21b

led principally to the " α -erythro" diastereomers 40a and 40c, respectively, favored both by "erythro-selectivity" and " α -selectivity" expectations. In the same fashion, the alkylation of methyl (3*R*)-3-ethyl-4-methylpentanoate (26) (entries 10 and 11, Table I) with 21a or 21b again led principally to the " α -erythro" diastereomers 27a and 27c, respectively, in which both "erythro-selectivity" and " α -selectivity" favored the same outcome. It was interesting that the level of diastereoselection at C-23 in these cases was not appreciably different from that observed in the absence of a C-24 stereocenter (*i.e.*, comparison of entries 6, 7, 10, and 11 with entries 1, 2, and 3, Table I). However, as shown in Table I, the alkylation of methyl (3*S*)-3,4-methylpentanoate (33a) (entries 8 and 9, Table I) with 21a or 21b favored the " β -erythro" diastereomers 40f or 40h over the " α -threo" diastereomers 40e and 40g. The dominance of "erythro-selectivity" over " α -selectivity" in entries 8 and 9 in Table I presumably resulted from the more pronounced influence of the proximal C-24 stereocenter than of the distal sterane stereocenters.

The syntheses of the C-23 and C-24 erythro-diastereomers of (20*R*)-5 α -dinosterane (2a) and (2d) were completed as shown in Scheme IV. The alkylation of methyl (3*S*)-3,4-dimethylpentanoate (33a) with 21a led to a 4:1 ratio of the inseparable erythro- and threo-diastereomers 40f and 40e. The reduction of this mixture provided the corresponding alcohols 41f and 41e that were readily separated by silica gel chromatography. An X-ray crystallographic study of the principal isomer, (20*R*,23*S*,24*R*)-5 α -dinosteran-29-ol (41f), confirmed the C-23*S* stereochemical assignment.²⁷ The further reduction of the mesylate of (20*R*,23*S*,24*R*)-41f provided (20*R*,23*S*,24*R*)-5 α -dinosterane (2a). As shown in Scheme IV, the repetition of the alkylation of 21a using methyl (3*R*)-3,4-dimethylpentanoate (33b), reduction of the C-23 epimeric esters 40a and 40b, separation of the principal alcohol 41a, and the further reduction of this alcohol led to (20*R*,23*R*,24*S*)-5 α -dinosterane (2d).

The alkylation of the saturated esters 33a and 33b with 21a in Scheme IV produced only minor amounts of the threo-esters 40e and 40b and was not a practical source of appreciable quantities of the threo-diastereomers of (20*R*)-5 α -dinosterane (2b) and (2c). As shown in Scheme V, the alkylation of the α,β -unsaturated ester, methyl 3,4-dimethyl-2-pentenoate (32), provided a selective route to these diastereomers. The alkylation of 32 with 21a provided a 2.4:1 ratio of the β,γ -unsaturated esters 39a and 39b. The reduction of the esters afforded the separable homoallylic alcohols 42a and 42b, respectively. Although the direct reduction of either of these homoallylic alcohols 42a and 42b to the corresponding saturated alcohols exhibited little diastereoselectivity at C-24, the conversion of the individual homoallylic alcohol 42a to the corresponding *tert*-butyldimethylsilyl ether, catalytic hydrogenation over platinum, and deprotection provided principally the saturated alcohol 41e and a lesser amount of 41a. In the same fashion, the homoallylic alcohol 42b provided principally 41b and a lesser amount of 41f. The final reduction of the mesylates of these threo-isomers 41e and 41b furnished the remaining biomarkers, (20*R*,23*R*,24*R*)-5 α -dinosterane (2b) and (20*R*,23*S*,24*S*)-5 α -dinosterane²⁸ (2c), respectively.

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(27) The details of the X-ray crystallographic study will be reported in *Acta Crystallogr., Sect. C*.

Table I. Diastereoselection in the Alkylation of Methyl (3*R*)-3-Ethyl-4-methylpentanoate (26), Methyl (2*r*)-3,4-Methyl-2-pentenoate (32), Methyl (3*S*)- or (3*R*)-3,4-Dimethylpentanoate (33a) and (33b), Methyl Pentanoate (35), and Methyl 4-Methylpentanoate (36)

entry	ester	electrophile	products and epimer ratios ^a	entry	ester	electrophile	products and epimer ratios ^a
1		21a	 5.1 : 1	7		21b	 4 : 1
2		21b	 5.3 : 1	8		21a	 1 : 4
3		21a	 6.6 : 1	9		21b	 1 : 4
4		21a	 2.4 : 1	10		21a	 3 : 1
5		21b	 2.2 : 1	11		21b	 3 : 1
6		21a	 4 : 1				

^a Product ratios were determined by GC analysis on an SE-30 fused silica gel capillary column (15 m) at 250–290 °C using He as a carrier gas.

In summary, the alkylations of various acyclic esters with chiral and achiral iodides (Table I and Figure 2) highlighted some of the factors that influence diastereoselection. The alkylation of the *saturated* esters, methyl (3*S*)- and (3*R*)-3,4-dimethylpentanoate (33a) and (33b), with (20*S*)-20-(iodomethyl)-4α-methyl-5α-pregnane (21a) provided predominantly the erythro-diastereomers, and the further reduction of these diastereomers provides (20*R*,23*S*,24*R*)- and (20*R*,23*R*,24*S*)-5α-dinosteranes (2a) and (2d). A similar alkylation of methyl (3*R*)-3-ethyl-4-methylpentanoate¹¹ (26) with 21a followed by decarbomethoxylation provided an avenue to (20*R*,24*R*)-4α-methyl-5α-stigmastane (1). The alkylation of the α,β-*unsaturated* ester methyl 3,4-dimethyl-2-pentenoate (32) with 21a and the further reduction of the products provided access to the three-diastereomers (20*R*,23*R*,24*R*)- and (20*R*,23*S*,24*S*)-5α-dinosterane (2b) and (2c). These approaches for the selective synthesis of individual diastereomers of sterane biomarkers will prove valuable in continuing studies to synthesize other biomarkers in the C₃₀ mono- and triaromatic dinosterane families.²⁹

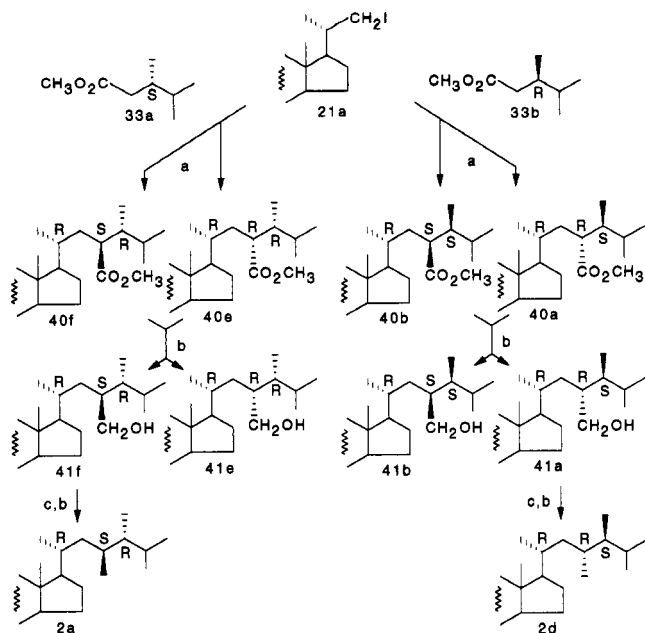
(28) It should be noted that the separation of 41b and 41f in Scheme V required a tedious HPLC separation. Consequently, the preparation of a pure sample of 2c was most easily accomplished using the minor isomer 41b that had been separated from the mixture of 41a and 41b produced by the route shown in Scheme IV.

Experimental Section

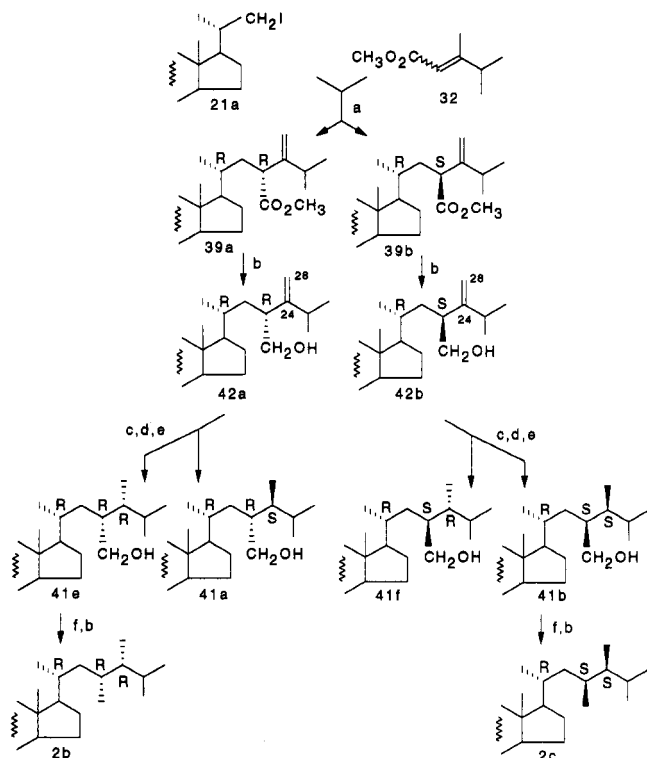
Pregnenolone Ethylene Ketal (11). The procedure of Bach^{12b} was repeated using 10 g (32 mmol) of pregnenolone (10) and 400 mg of *p*-toluenesulfonic acid monohydrate in 350 mL of distilled ethylene glycol to afford, after the ethylene glycol was distilled from the mixture at 80–85 °C (1 mm) over a 9-h period, 9 g (79%) of 11: mp 158–61 °C (lit.^{12b} mp 160–163 °C); ¹H NMR (CDCl₃) δ 0.70 (s, 3, C-18 CH₃), 1.01 (s, 3, C-19 CH₃), 1.30 (s, 3, C-21 CH₃), 3.40–3.60 (m, 1, CH(OH)), 3.80–4.08 (m, 4, OCH₂-CH₂O), 5.34–5.40 (m, 1, C-6 vinylic H); ¹³C NMR (CDCl₃) δ 12.6, 19.2, 20.6, 22.7, 23.6, 24.4, 31.2, 31.4, 31.6, 36.3, 37.1, 39.2, 41.6, 42.1, 49.9, 56.5, 58.1, 63.1, 65.1, 71.7, 112.1, 121.7, 141.0.

Progesterone 20-Ethylene Ketal (12). The procedure of Raggio¹³ was repeated using 18.5 g (51.4 mmol) of 11 in 300 mL of toluene, 48 mL (44.1 g, 7.6 equiv) of distilled 1-methyl-4-piperidone, and 30.5 g (0.15 mol, 2.91 equiv) of Al(*O*-*i*-Pr)₃ under a Dean-Stark trap to afford, after chromatography on a silica gel column using 1:1 EtOAc-hexane, 14.3 g (78%) of 12, mp 189.5–190.5 °C. A sample was recrystallized from MeOH: mp 190–192 °C (lit.^{11a} mp 189–90 °C; lit.^{11b} mp 187–192.5 °C); ¹H NMR (CDCl₃) δ 0.81 (s, 3, C-18 CH₃), 1.19 (s, 3, C-19 CH₃), 1.30 (s, 3, C-21 CH₃),

(29) (a) Lichtfouse, E.; Riolo, J.; Albrecht, P. *Tetrahedron Lett.* 1990, 31, 3937. (b) Carlson, R. M. K.; Chamberlain, D. E. *Org. Geochem.* 1986, 10, 163. (c) Shi, J. Y.; Mackenzie, A. S.; Alexander, R.; Eglinton, G.; Gowar, A. P.; Wolff, G. A.; Maxwell, J. R. *Chem. Geol.* 1982, 35, 1. (d) Ludwig, B.; Hussler, G.; Wehrung, P.; Albrecht, P. *Tetrahedron Lett.* 1981, 22, 3313. (e) Mackenzie, A. S.; Hoffmann, C. F.; Maxwell, J. R. *Geochim. Cosmochim. Acta* 1981, 45, 1345.

Scheme IV. Synthesis of Erythro-Diastereomers of 5 α -Dinosterane (2)^a


^a Key: (a) LDA, HMPA, THF, -78 °C; (b) LiAlH₄ followed by chromatographic separation; (c) MsCl, Et₃N.

Scheme V. Synthesis of Threo-Diastereomers of 5 α -Dinosterane (2)^a


^a Key: (a) LDA, HMPA, THF, -78 °C; (b) LiAlH₄; (c) TBSCl, imidazole; (d) H₂, PtO₂; (e) (n-Bu)₄NF; (f) MsCl, Et₃N.

3.80–4.10 (m, 4, OCH₂CH₂O), 5.72–5.78 (m, 1, C-6 vinylic H); ¹³C NMR (CDCl₃) δ 12.6, 17.1, 20.5, 22.6, 23.5, 24.3, 31.7, 32.7, 33.8, 34.9, 35.5, 38.4, 39.1, 41.6, 53.6, 55.6, 58.0, 63.1, 65.1, 111.9, 123.9, 171.9, 200.1.

4-((Phenylthio)methyl)-4-pregnene-3,20-dione 20-Ethylene Ketal (13). The procedure of Kirk¹⁴ was repeated with modifications. A mixture of 14.6 g (40.7 mmol) of 12, 11.7 g (10.9 mL, 2.6 equiv) of thiophenol, 8.75 g (21.8 mL, 7.16 equiv) of a 37% aqueous solution of formaldehyde, and 8.73 g (12 mL, 2.12

equiv) of Et₃N in 90 mL of EtOH was refluxed for 72 h. The EtOH was evaporated; the yellow viscous oil was dissolved in Et₂O; and the organic solution was washed successively with aqueous NaOH solution and brine and dried over anhydrous MgSO₄. The solvents were evaporated to afford a yellow viscous oil which was chromatographed on a silica gel column using 1:5 EtOAc-hexane to give 10.6 g (54%) of 13: mp 124–124.5 °C; IR (CHCl₃) 1660 (C=O), 1595 (C=C) cm⁻¹; ¹H NMR (CDCl₃) δ 0.79 (s, 3, C-18 CH₃), 1.15 (s, 3, C-19 CH₃), 1.29 (s, 3, C-21 CH₃), 3.82–4.04 (m, 6, OCH₂CH₂O and CH₂SPh), 7.17–7.43 (m, 5, ArH); ¹³C NMR (CDCl₃) δ 12.6, 17.6, 20.5, 22.7, 23.4, 24.3, 27.9, 28.6, 31.6, 33.4, 34.4, 34.6, 39.1, 39.2, 41.6, 54.0, 55.6, 58.0, 63.1, 65.1, 111.8, 126.6, 128.3, 128.8, 131.1, 136.6, 168.5, 197.4 (C=O); exact mass spectrum calcd for C₃₀H₄₀SO₃ 480.2700, found 480.2699. Anal. Calcd for C₃₀H₄₀SO₃: C, 74.95; H, 8.39. Found: C, 74.84; H, 8.42.

4 α -Methyl-5 α -pregnane-3,20-dione 20-Ethylene Ketal (14). To 1.63 g (235 mmol, 3.7 equiv) of lithium in 250 mL of liquid NH₃ (distilled from sodium) at -78 °C under N₂ was added 5.07 g (10.5 mmol) of 13 in 50 mL of anhydrous THF. The dark blue solution was stirred at -78 °C for 5 h, and the reaction was quenched with 15 g of NH₄Cl. To this solution was added 100 mL of Et₂O, and the NH₃ was allowed to evaporate. The residue was dissolved in water and extracted with Et₂O. The organic layer was washed with brine, dried over anhydrous MgSO₄, concentrated, and chromatographed on silica gel using 1:5 EtOAc-hexane to furnish 2.15 g (54%) of 14: mp 200–202 °C; IR (CHCl₃) 1695 (C=O) cm⁻¹; ¹H NMR (CDCl₃) δ 0.78 (s, 3, C-18 CH₃), 0.97 (d, *J* = 6.6 Hz, 3, C-4 α CH₃), 1.07 (s, 3, C-19 CH₃), 1.29 (s, 3, C-21 CH₃), 3.82–4.10 (m, 4, OCH₂CH₂O); ¹³C NMR (CDCl₃) δ 11.2, 12.5, 12.8, 20.9, 22.7, 23.5, 24.4, 25.4, 31.6, 34.2, 36.2, 37.9, 39.1, 39.4, 41.8, 44.9, 53.5, 53.9, 56.1, 58.2, 63.1, 65.1, 112.0, 214.3 (C=O). Anal. Calcd for C₂₄H₃₈O₃: C, 76.96; H, 10.23. Found: C, 76.72; H, 10.25.

4 α -Methyl-3 β -hydroxy-5 α -pregnan-20-one Ethylene Ketal (15). To 125 mg (3.29 mmol, 2 equiv) of LiAlH₄ in 50 mL of anhydrous THF was added 2.47 g (6.59 mmol) of 14 in 115 mL of anhydrous THF dropwise over a period of 30 min. The mixture was stirred at 25 °C for 2.5 h. To the mixture at 0 °C was added slowly 125 mL of water followed by 375 mL of 15% aqueous NaOH and finally by 375 mL of water. The product was diluted with Et₂O, filtered, and washed with brine. The Et₂O solution was dried over anhydrous MgSO₄ and concentrated to afford 2.15 g (87%) of 15 that was principally the C-3 β epimer. An analytical sample was prepared by recrystallization from MeOH: mp 193–195.5 °C; IR (CHCl₃) 3610 (OH) cm⁻¹; ¹H NMR (CDCl₃) δ 0.75 (s, 3, C-18 CH₃), 0.83 (s, 3, C-19 CH₃), 0.95 (d, *J* = 6.2 Hz, 3, C-4 α CH₃), 1.29 (s, 3, C-21 CH₃), 3.00–3.20 (m, 1, CHOH), 3.80–4.05 (m, 4, CH₂CH₂O); ¹³C NMR (CDCl₃) δ 12.8, 13.1, 14.9, 20.7, 22.7, 23.5, 23.9, 24.4, 30.9, 31.9, 34.1, 35.8, 39.0, 39.5, 41.8, 50.8, 54.4, 56.3, 58.2, 63.1, 65.2, 76.5, 112.1. Anal. Calcd for C₂₄H₄₀O₃: C, 76.54; H, 10.70. Found: C, 76.38; H, 10.65.

4 α -Methyl-3 β -hydroxy-5 α -pregnan-20-one Ethylene Ketal Phenyl Thiocarbonate (16). The procedure of Robins^{16c} was repeated using 408 mg (1.08 mmol) of 15, 281 mg (225 mL, 1.62 mmol) of phenyl thionochloroformate, 343 mg (347 μ L, 4.33 mmol) of anhydrous pyridine, and 24 mL of anhydrous CH₂Cl₂ to afford, after chromatography on silica gel using 1:10 EtOAc-hexane, 401 mg (78%) of 16: mp 195–196.5 °C; IR (CHCl₃) 1550, 1470, 1430, 1365, 1355, 1315, 1275, 1175, 1130, 1055, 1035 cm⁻¹; ¹H NMR (CDCl₃) δ 0.76 (s, 3, C-18 CH₃), 0.88 (s, 3, C-19 CH₃), 0.94 (d, *J* = 6.2 Hz, 3, C-4 α CH₃), 1.29 (s, 3, C-21 CH₃), 3.80–4.05 (m, 4, OCH₂CH₂O), 4.82–5.0 (m, 1, CHOC(S)OPh), 7.0–7.5 (m, 5, Ar); ¹³C NMR (CDCl₃) δ 12.8, 13.0, 14.9, 20.7, 22.7, 23.4, 23.8, 24.3, 25.6, 31.8, 34.1, 35.7, 36.0, 36.1, 39.4, 41.7, 50.8, 54.1, 56.2, 58.2, 63.1, 65.1, 90.1, 112.0, 122.2, 126.5, 129.6, 153.5, 195.3. Anal. Calcd for C₃₁H₄₄SO₃: C, 72.61; H, 8.65. Found: C, 72.55; H, 8.68.

4 α -Methyl-5 α -pregnan-20-one Ethylene Ketal (17). A solution of 3.05 g (2.83 mL, 10.5 mmol) of (n-Bu)₃SnH in 30 mL of anhydrous benzene was added dropwise over a 3.5-h period to a refluxing solution of 1.63 g (3.18 mmol) of 16 and 0.052 g (0.32 mmol) of 2,2'-azobisisobutyronitrile in 200 mL of anhydrous benzene. The mixture was refluxed an additional 4 h, cooled, and diluted with Et₂O. The solution was washed successively with cold 1 M HCl solution, water, 5% NaOH, and brine, and dried over anhydrous MgSO₄. The crude product was chro-

matographed on silica gel using 1:10 EtOAc-hexane to afford 945 mg (82%) of **17**: mp 134–135 °C; IR (CHCl₃) 3000–2820, 1430, 1370, 1230–1190, 1040 cm⁻¹; ¹H NMR (CDCl₃) δ 0.74 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6 Hz, 3, C-4α CH₃), 1.29 (s, 3, C-21 CH₃), 3.82–4.10 (m, 4, OCH₂CH₂O); ¹³C NMR (CDCl₃) δ 12.8, 13.0, 20.4, 20.6, 21.7, 22.7, 23.5, 24.1, 24.3, 31.1, 32.0, 34.2, 36.3, 36.4, 38.7, 39.6, 41.7, 53.4, 54.7, 56.5, 58.3, 63.1, 65.2, 112.1 (C-20). Anal. Calcd for C₂₄H₄₀O₂: C, 79.94; H, 11.18. Found: C, 79.98; H, 11.14.

4α-Methyl-5α-pregnan-20-one (18). A solution of 1.01 g (2.8 mmol) of **17** in 120 mL of 1:2.5:5.5 mixture of 1 M HCl-HOAc-THF was stirred at 25 °C for 2 h. The mixture was poured into 200 mL of cold water. The organic was separated, and the aqueous layer was extracted with EtOAc. The combined organic layers were washed successively with saturated NaHCO₃ solution and brine and dried over anhydrous MgSO₄. The product was chromatographed on silica gel using 1:10 EtOAc-hexane to afford 760 mg (86%) of **18**: mp 159–160 °C; IR (CHCl₃) 1705 (C=O) cm⁻¹; ¹H NMR (CDCl₃) δ 0.59 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.80 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 2.11 (s, 3, C-21 CH₃); ¹³C NMR (CDCl₃) δ 13.0, 13.2, 20.3, 20.8, 21.6, 22.5, 23.9, 24.1, 31.0, 31.3, 32.0, 34.9, 36.3, 38.7, 39.0, 44.0, 53.3, 54.5, 56.8, 63.8, 210.2 (21 lines; one line represents two carbons); ¹³C NMR (CDCl₃) (with 5% of Eu(fod)₃) δ 13.3, 13.9, 20.6, 21.1, 21.9, 23.3, 24.2, 24.6, 31.3, 32.3, 32.7, 35.2, 36.49, 36.54, 38.9, 39.5, 44.7, 53.5, 54.8, 57.2, 64.5, 211.9; exact mass spectrum calcd for C₂₂H₃₆O 316.2767, found 316.2766. Anal. Calcd for C₂₂H₃₆O: C, 83.48; H, 11.47. Found: C, 83.40; H, 11.44.

4α-Methyl-20-methylene-5α-pregnane (19). To a suspension of 2.56 g (7.17 mmol) of methyltriphenylphosphonium bromide in 8 mL of anhydrous benzene under N₂ was added 905 mg of 1.66 M (7.17 mmol) potassium *tert*-amylate in benzene. The mixture was refluxed for 1 h, and 337 mg (1.06 mmol) of **18** in 7.5 mL of anhydrous benzene was added. The mixture was refluxed for 4.5 h and stirred for 12 h at 25 °C. The mixture was poured into water and extracted with Et₂O. The combined organic layers were washed successively with 10% HCl solution, saturated NaHCO₃ solution, and brine, and dried over anhydrous MgSO₄. The crude product was chromatographed twice on silica gel with hexane to afford 306 mg (91%) of **19**: mp 156.5–159 °C; IR (CHCl₃) 1630 (C=C), 1440, 1370 cm⁻¹; ¹H NMR (CDCl₃) δ 0.55 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.80 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 1.75 (s, 3, C-21 CH₃), 4.70 (s, 1, vinylic H), 4.84 (s, 1, vinylic H); ¹³C NMR (CDCl₃) δ 12.6, 13.0, 20.4, 20.8, 21.7, 23.9, 24.1, 24.5, 25.2, 31.1, 32.0, 35.3, 36.4, 38.7, 38.9, 43.1, 53.4, 54.8, 56.4, 57.3, 110.6 (C-22), 146.1 (C-20). Anal. Calcd for C₂₈H₃₈: C, 87.82; H, 12.18. Found: C, 87.79; H, 12.15.

(20S)- and (20R)-20-(Hydroxymethyl)-4α-methyl-5α-pregnane (20a) and (20b) from 19. To a suspension of 362 mg (1.15 mmol) of **19** in 1.15 mL of anhydrous THF at 0 °C under N₂ was added 864 mL (0.864 mmol) of 1 M borane-THF solution via a syringe over a 10-min period. The ice bath was removed, and the clear solution was stirred at 25 °C for 1 h. The mixture was again cooled to 0 °C, and the excess borane was destroyed with 100 mL of water. To this solution was added 960 mL (2.88 mmol) of 3 M NaOH solution followed by 960 mL (2.88 mmol) of a 30% H₂O₂ solution. The mixture was initially viscous and was heated for 3.5 h at 53–54 °C. The solution was cooled, and the phases were separated. The aqueous layer was extracted with Et₂O. The combined organic layers were washed with brine and dried over anhydrous MgSO₄. The product was chromatographed on silica gel using 1:5 EtOAc-hexane to afford 312 mg (81%) of **20** as a C-20 epimeric mixture: mp 157–159 °C; IR (CHCl₃) 3610 (OH), 1445, 1435, 1370 cm⁻¹; ¹H NMR (CDCl₃) δ 0.67 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.80 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 0.95 and 1.03 (two d, *J* = 6.6 Hz, 3, C-21S and C-21R CH₃), 3.30–3.80 (m, 2, CH₂OH). Anal. Calcd for C₂₃H₄₀O: C, 83.06; H, 12.12. Found: C, 82.94; H, 12.09. The mixture was separated on silica gel using 1:10 EtOAc-hexane to afford the individual epimers having the following physical and spectral data. Data for the C-20S epimer **20a**: mp 156.5–157 °C; IR (CHCl₃) 3610 (OH), 1445, 1435, 1370 cm⁻¹; ¹H NMR (CDCl₃) δ 0.67 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 0.95 (d, *J* = 6.6 Hz, 3, C-21 CH₃), 3.40–3.80 (m, 2, CH₂OH); ¹³C NMR (CDCl₃) δ 12.2, 13.0, 16.5, 20.4, 20.7, 21.7, 23.8, 24.0, 27.5, 31.1, 32.0, 34.8, 36.2, 36.3, 37.8, 38.7, 39.5, 42.1, 54.2, 53.3, 54.6, 56.5,

66.8 (CH₂OH). Data for the C-20R epimer **20b**: mp 167–168 °C; IR (CHCl₃) 3610 (OH), 1435, 1370 cm⁻¹; ¹H NMR (CDCl₃) δ 0.67 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.80 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 1.03 (d, *J* = 6.6 Hz, 3, C-21 CH₃), 3.30–3.70 (m, 2, CH₂OH); ¹³C NMR (CDCl₃) δ 11.9, 13.0, 16.4, 20.4, 20.7, 21.7, 24.0, 27.6, 31.1, 32.0, 34.9, 36.2, 36.3, 38.7, 39.8, 42.4, 52.4, 53.3, 54.6, 56.3, 68.0 (CH₂OH).

(20S)- and (20R)-20-(Hydroxymethyl)-4α-methyl-5α-pregnane (20a) and (20b) from 25. The procedure of Shu^{6c} was repeated using 410 mg (1 mmol) of **25** in 20 mL of CH₂Cl₂ and a slight excess of O₃. The mixture was quenched with 369 μL (5 mmol, 5 equiv) of dimethyl sulfide and warmed to 25 °C. The solution was concentrated and dissolved in 10 mL of MeOH containing 40 mg (1 mmol, 1 equiv) of NaOH. The solution was refluxed for 5 min and cooled. To this solution was added 76 mg (2 mmol, 8 equiv) of NaBH₄ to afford 241 mg (73%) of **20a** and **20b** as a 1:1 mixture. In the absence of the methanolic NaOH equilibration step, **25** was converted exclusively to **20a**.

(20S)-20-(Iodomethyl)-4α-methyl-5α-pregnane (21a). The procedure of Crossland¹⁸ was repeated using 1.33 g (4 mmol) of **20a** and 504 mg (4.4 mmol, 1.1 equiv) of CH₃SO₂Cl to give 1.57 g (96%) of (20S)-20-hydroxymethyl-4α-methyl-5α-pregnane mesylate. The procedure of Partridge¹⁹ was repeated using 1.57 g (3.8 mmol) of (20S)-20-(hydroxymethyl)-4α-methyl-5α-pregnane mesylate and 5.76 (38 mmol, 10 equiv) of NaI to give 1.44 g (85%) of **21a**: mp 152–153 °C; IR (CHCl₃) 2910, 1440, 1370 cm⁻¹; ¹H NMR (CDCl₃) δ 0.68 (s, 3, C-18 CH₃), 0.79 (d, *J* = 6.2 Hz, 3, C-4α CH₃), 0.79 (s, 3, C-19 CH₃), 1.01 (d, *J* = 5.2 Hz, 3, C-21 CH₃), 3.12–3.36 (m, 2, CH₂I); ¹³C NMR (CDCl₃) δ 12.8, 13.2, 20.6, 20.7, 20.9, 21.3, 21.9, 24.0, 24.2, 27.6, 31.2, 32.2, 35.1, 36.4, 36.5, 37.0, 38.8, 39.8, 42.5, 53.4, 54.6, 55.5, 56.3. Anal. Calcd for C₂₃H₃₉I: C, 62.43; H, 8.88. Found: C, 62.41; H, 8.91.

(20R)-20-(Iodomethyl)-4α-methyl-5α-pregnane (21b). The procedure of Crossland¹⁸ was repeated using 253 mg (0.76 mmol) of **20b** and 96 mg (0.84 mmol, 1.1 equiv) of CH₃SO₂Cl to give 277 mg (89%) of (20R)-20-(hydroxymethyl)-4α-methyl-5α-pregnane mesylate. The procedure of Partridge¹⁹ was repeated using 277 mg (0.68 mmol) of (20R)-20-(hydroxymethyl)-4α-methyl-5α-pregnane mesylate and 977 mg (6.7 mmol, 10 equiv) of NaI to give 246 mg (84%) of **21b**: mp 136.5–138 °C; IR (CHCl₃) 2900, 1430, 1360 cm⁻¹; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.79 (d, *J* = 6.2 Hz, 3, C-4α CH₃), 0.96 (d, *J* = 6 Hz, 3, C-21 CH₃), 3.20–3.50 (m, 2, CH₂I); ¹³C NMR (CDCl₃) δ 12.2 (C-18), 13.0 (C-19), 19.2, 20.4, 20.7, 21.3, 21.6, 23.7, 24.0, 27.7, 31.0, 32.0, 34.9, 36.3, 36.4, 36.6, 38.7, 39.8, 42.2, 53.3, 54.4, 54.6, 56.2. Anal. Calcd for C₂₃H₃₉I: C, 62.43; H, 8.88. Found: C, 62.51; H, 8.91.

(20R,22E,24R)-4-((Phenylthio)methyl)-4,22-stigmastadien-3-one (23). The procedure described for the preparation of **13** was repeated using 16.7 g (40.7 mmol) of (22E)-stigmastadien-3-one,¹³ 1.66 g (10.9 mL, 105 mmol, 2.6 equiv) of thiophenol, 88 g (21.8 mL, 292 mmol, 7.2 equiv) of 37% formalin, and 8.7 g (12 mL, 86.3 mmol, 2.1 equiv) of Et₃N in 90 mL of EtOH at reflux for 72 h to afford an oil which crystallized upon storing at -5 °C for 2–3 days. The solid was recrystallized from 1:3 Et₂O-EtOH followed by a second recrystallization from 1:2 Et₂O-EtOH to yield 13.4 g (62%) of **23**: mp 92–93 °C; IR (CHCl₃) 1660 (C=O), 1595 (C=C) cm⁻¹; ¹H NMR (CDCl₃) δ 0.71 (s, 3, C-18 CH₃), 0.78–0.86 (m, 9, C-26, C-27, C-29 CH₃), 1.01 (d, *J* = 6.4 Hz, 3, C-21 CH₃), 1.14 (s, 3, C-19 CH₃), 3.87 (s, 2, CH₂SPh), 4.95–5.21 (m, 2, CH=CH), 7.18–7.40 (m, 5, ArH); ¹³C NMR (CDCl₃) δ 11.8 (C-18), 11.9 (C-19), 17.5, 18.7, 20.7, 20.8 (two C), 23.8, 25.1, 27.9, 28.5, 28.6, 31.6, 31.7, 33.4, 34.5, 34.8, 39.1, 39.3, 40.2, 42.0, 51.0, 53.9, 55.6, 55.7, 126.5, 128.2, 128.7, 129.4, 131.0 (C-23), 136.6 (C-22), 138.2 (C-4), 168.4 (C-5), 197.2 (C=O). Anal. Calcd for C₃₆H₅₂SO: C, 81.14; H, 9.84. Found: C, 81.00; H, 9.86.

(20R,22E,24R)-4α-Methyl-5α-stigmast-22-en-3β-ol (24). The procedure described for the preparation of **14** was repeated using 7.9 g (14.8 mmol, 1 equiv) of **23** and 2.5 g (357 mmol, 3 equiv) of Li in 300 mL of NH₃ and using EtOH instead of NH₄Cl during the quenching process to afford 5.2 g (82%) of **24**: mp 92–93 °C; IR (CHCl₃) 3610 (OH), 980 cm⁻¹; ¹H NMR (CDCl₃) δ 0.67 (s, 3, C-18 CH₃), 0.79 (d, *J* = 6.4 Hz, 3, C-26 CH₃), 0.80 (t, *J* = 7 Hz, 3, C-29 CH₃), 0.83 (s, 3, C-19 CH₃), 0.84 (d, *J* = 6.4 Hz, 3, C-27 CH₃), 0.94 (d, *J* = 6.2 Hz, 3, C-4α CH₃), 1.00 (d, *J* = 6.4 Hz, 3, C-21 CH₃), 3.02–3.14 (m, 1, CHOH), 4.94–5.21 (m, 2, CH=CH);

^{13}C NMR (CDCl_3) δ 11.99 (C-18), 12.02 (C-19), 13.1, 14.9, 18.8, 20.9, 21.0, 23.96, 24.03, 25.2, 28.8, 30.8, 31.7, 32.0, 34.6, 35.8, 36.6, 39.0, 39.8, 40.4, 42.2, 50.8, 51.1, 54.4, 55.9, 56.5, 76.5 (CHOH), 128.7 (C-23), 138.0 (C-22). Anal. Calcd for $\text{C}_{30}\text{H}_{52}\text{O}$: C, 84.04; H, 12.23. Found: C, 84.11; H, 12.21.

(20R,22E,24R)-4 α -Methyl-5 α -stigmast-22-ene (25). The procedure of Robins^{16c} was repeated using 12.5 g (29 mmol) of **24**, 6 g (35 mmol, 1.2 equiv) of phenyl thionochloroformate, and 11 g (38 mmol, 1.3 equiv) of $n\text{-Bu}_3\text{SnH}$ to afford, after chromatography on silica gel using hexane and recrystallization from 1:1 EtOAc–EtOH, 9.8 g (81%) of **25**: mp 119–120 °C; ^1H NMR (CDCl_3) δ 0.66 (s, 3, C-18 CH_3), 0.75–0.92 (m, 15, C-4 α , C-19, C-26, C-27, C-29 CH_3), 1.01 (d, $J = 6.6$ Hz, 3, C-21 CH_3), 4.93–5.22 (m, 2, $\text{CH}=\text{CH}$); ^{13}C NMR (CDCl_3) δ 12.7 (C-19), 13.7 (C-18), 19.4, 21.0, 21.4, 21.5, 21.6, 22.3, 23.1, 24.7, 25.9, 29.5, 30.2, 31.8, 32.4, 32.7, 35.6, 36.9, 37.0, 39.4, 40.6, 41.1, 42.8, 51.8, 54.0, 55.4, 56.6, 57.4, 129.9 (C-23), 139.3 (C-22). Anal. Calcd for $\text{C}_{30}\text{H}_{52}$: C, 87.30; H, 12.70. Found: C, 87.32; H, 12.67.

Methyl (20R,23 β ,24S)-4 α -Methyl-5 α -stigmastane-23-carboxylate (27). The procedure described for the preparation of **40e** and **40f** (vide infra) was repeated using 44 mg (0.1 mmol) of **21a** with 79 mg (0.5 mmol) of methyl (3R)-3-ethyl-4-methyl-pentanoate¹¹ (**26**) to afford 37 mg (78%) of **27a** and **27b** as a mixture: IR (CHCl_3) 1730 (C=O), 1460, 1430, 1360, 1165 cm^{-1} ; ^1H NMR (CDCl_3) δ 0.62 and 0.64 (two s, 3, C-18 CH_3), 0.75–0.94 (m, 18, C-4 α , C-19, C-21, C-26, C-27, C-29 CH_3), 2.48–2.60 (m, 1, CHCO_2CH_3); exact mass spectrum calcd for $\text{C}_{32}\text{H}_{54}\text{O}_2$ 472.4283, found 472.4285. GC analysis on an SE-30 fused silica gel capillary column (15 m) indicated that the product was a 3:1 ratio of **27a** and **27b**.

(20R,23 β ,24S)-4 α -Methyl-5 α -stigmastane-23-carboxylic Acid (28). To 35 mg (0.074 mmol) of **27** in 1 mL of collidine was added 99 mg (0.74 mmol, 10 equiv) of anhydrous LiI. The mixture was refluxed at 172 °C under N_2 for 12 h, cooled, diluted with 2 M HCl solution, and extracted with Et_2O . The combined organic solutions were washed with 10% $\text{Na}_2\text{S}_2\text{O}_3$ solution and dried over anhydrous MgSO_4 . The product was chromatographed on silica gel using 1:10 EtOAc–hexane to afford 28 mg (82%) of **28**: ^1H NMR (CDCl_3) δ 0.64 (s, 3, C-18 CH_3), 0.72–0.92 (m, 18, C-4 α , C-19, C-21, C-26, C-27, C-29 CH_3), 2.35–2.55 (m, 1, CHCO_2H); exact mass spectrum calcd for $\text{C}_{31}\text{H}_{52}\text{O}$ 458.4124, found 458.4129.

Phenylselenenyl (20R,23 β ,24S)-4 α -Methyl-5 α -stigmastane-23-carboxylate (29). To a solution of 25 mg (0.054 mmol, 1 equiv) of **28** in 0.5 mL of anhydrous THF at 0 °C under N_2 was added 23 μL (16 mg, 0.162 mmol, 3 equiv) of anhydrous Et_3N and 16.1 μL (23 mg, 0.108 mmol, 2 equiv) of $\text{PhOP}(\text{O})\text{Cl}_2$. After 30 min, an additional 37.5 μL (27 mmol, 5 equiv) of anhydrous Et_3N and 22.9 μL (34 mg, 0.216 mmol, 4 equiv) of benzeneselenol were added. After 30 min, the mixture was allowed to warm to 25 °C. The product was diluted with 1:1 hexane– Et_2O , washed twice with water and brine, and dried over anhydrous MgSO_4 to give a yellow oil that solidified at –5 °C. The crude selenoester was purified by column chromatography on silica gel using hexane followed by 5% EtOAc–hexane to afford 23 mg (69%) of **29**. This material was used directly in the next step without further purification.

(20R,24R)-4 α -Methyl-5 α -stigmastane (1) from 29. To a refluxing solution of 23 mg (0.038 mmol, 1 equiv) of **29** and 51 μL (55 mg, 0.91 mmol, 5 equiv) of $(n\text{-Bu})_3\text{SnH}$ in 2 mL of anhydrous toluene under N_2 was added 0.6 mg (3.8 μmol , 0.1 equiv) of 2,2'-azobisisobutyronitrile. The solution was refluxed for 2 h, cooled, and concentrated. The product was treated with O_3 in 2 mL of hexane for 5 min at –78 °C. The solution was warmed to 25 °C and passed through a silica gel column. The hydrocarbon was eluted with hexane to afford 10 mg (62%) of **1** that was identical in all respects with **1** obtained from (22E)-stigmasterol (**22**) (vide infra).

(20R,24R)-4 α -Methyl-5 α -stigmastan-3 β -ol (30). A mixture of 5 g (11.7 mmol) of **24** and 500 mg of 10% Pd–C in 150 mL of anhydrous benzene was hydrogenated at 60 psi and 25 °C for 48 h to afford 4.8 g (96%) of **30**: mp 184–185 °C; IR (CHCl_3) 3430 (OH) cm^{-1} ; ^1H NMR (CDCl_3) δ 0.65 (s, 3, C-18 CH_3), 0.81 (d, $J = 6.8$ Hz, 3, C-26 CH_3), 0.83 (s, 3, C-19 CH_3), 0.83 (d, $J = 6.8$ Hz, 3, C-27 CH_3), 0.84 (t, $J = 7.2$ Hz, 3, C-29 CH_3), 0.90 (d, $J = 6.6$ Hz, 3, C-21 CH_3), 0.95 (d, $J = 6.4$ Hz, 3, C-4 α CH_3); ^{13}C NMR

(CDCl_3) δ 11.7 (C-18), 11.8 (C-19), 13.1 (C-4), 14.9, 18.5, 18.8, 19.6, 20.9, 22.8, 24.0 (two C), 25.8, 28.1, 28.9, 30.9, 32.0, 33.7, 34.6, 35.8, 36.0, 36.6, 39.0, 39.9, 42.3, 45.6, 50.8, 54.4, 56.4, 76.5 (CHOH). Anal. Calcd for $\text{C}_{30}\text{H}_{54}\text{O}$: C, 83.65; H, 12.64. Found: C, 83.68; H, 12.82.

(20R,24R)-4 α -Methyl-5 α -stigmastane (1) from 30. The procedure of Robins^{16c} was repeated using 4.3 g (10 mmol, 1 equiv) of **30** and 2.6 g (2.1 mL, 15 mmol, 1.5 equiv) of phenyl chlorothionochloroformate to afford 4.4 g (78%) of (20R,24R)-4 α -methyl-5 α -stigmastan-3 β -ol phenyl thiocarbonate: mp 181–181.5 °C (from 2:1 EtOAc–EtOH); IR (CHCl_3) 1560, 1470, 1360 cm^{-1} ; ^1H NMR (CDCl_3) δ 0.66 (s, 3, C-18 CH_3), 0.81 (d, $J = 6.8$ Hz, 3, C-26 CH_3), 0.83 (d, $J = 6.8$ Hz, 3, C-27 CH_3), 0.84 (t, $J = 6.8$ Hz, 3, C-29 CH_3), 0.88 (s, 3, C-19 CH_3), 0.91 (d, $J = 7.2$ Hz, 3, C-21 CH_3), 0.94 (d, $J = 6.6$ Hz, 3, C-4 α CH_3), 4.85–4.98 (m, 1, $\text{CHOC}(\text{S})\text{OPh}$); ^{13}C NMR (CDCl_3) δ 11.7 (C-18), 11.8 (C-19), 13.0 (C-4), 15.0, 18.5, 18.8, 19.6, 20.9, 22.8, 23.9, 24.0, 25.6, 25.8, 28.1, 28.9, 31.9, 33.7, 34.6, 35.7, 36.0, 36.1, 36.2, 39.8, 42.3, 45.6, 50.8, 54.2, 56.0, 56.3, 90.2 ($\text{CHOC}(\text{S})\text{OPh}$), 122.2 (two C), 126.6, 129.6 (two C), 153.6, 195.3 (C=S). Anal. Calcd for $\text{C}_{37}\text{H}_{58}\text{SO}_2$: C, 78.39; H, 10.31. Found: C, 78.42; H, 10.27. The procedure of Robins^{16c} was repeated using 4.3 g (7.7 mmol, 1 equiv) of (20R,24R)-4 α -methyl-5 α -stigmastan-3 β -ol phenyl thiocarbonate and 3.4 g (3.1 mL, 11.6 mmol, 1.5 equiv) of $(n\text{-Bu})_3\text{SnH}$ to afford 2.6 g (83%) of **1**: mp 92.5–93.5 °C (from 2:1 EtOAc–EtOH); IR (CHCl_3) 1450, 1440, 1360 cm^{-1} ; ^1H NMR (CDCl_3) δ 0.65 (s, 3, C-18 CH_3), 0.79 (s, 3, C-19 CH_3), 0.80–0.88 (m, 12, C-4 α , C-25, C-26, C-27 CH_3), 0.91 (d, $J = 6.4$ Hz, 3, C-21 CH_3); ^{13}C NMR (CDCl_3) δ 11.7 (C-18), 11.8 (C-19), 13.0 (C-4), 18.5, 18.8, 19.6, 20.4, 20.8, 21.7, 22.9, 24.0, 24.1, 25.9, 28.2, 28.9, 31.1, 32.1, 33.8, 34.9, 36.1, 36.3, 36.4, 38.7, 40.0, 42.0, 45.7, 53.4, 54.7, 56.1, 56.6. Anal. Calcd for $\text{C}_{30}\text{H}_{54}$: C, 86.88; H, 13.12. Found: C, 86.80; H, 13.04.

Methyl (2E)- and (2Z)-3,4-Dimethyl-2-pentenoate (32a) and (32b). The procedure of Huffman²⁵ was repeated using 4.32 g (109 mmol) of 60% NaOH solution, 20 g (108.5 mmol) of methyl dimethylphosphonoacetate, and 9.27 g (110 mmol) of 3-methyl-2-butanone in 290 mL of anhydrous 1,2-dimethoxyethane under N_2 to afford, after heating at 60 °C for 66 h, 8.83 g (58%) of **32** as a mixture of *E* and *Z* isomers: bp 105–108 °C (92 mm); IR (CHCl_3) 1730 (C=O), 1705 (C=C), 1465 cm^{-1} ; ^1H NMR (CDCl_3) δ 1.00–1.12 (m, 6, $\text{CH}(\text{CH}_3)_2$), 1.80 and 2.14 (two d, $J = 1$ Hz, 3, CHCH_3 , *E* and *Z* isomers), 2.25–2.50 (m, 1, $\text{CH}(\text{CH}_3)_2$), 3.67 and 3.69 (two s, 3, OCH_3 , *E* and *Z* isomers), 5.58–5.62 and 5.67–5.71 (two m, 1, *E* and *Z* vinylic H); ^{13}C NMR (CDCl_3) δ 16.1, 19.0, 20.3, 20.6, 28.8, 37.9, 50.6, 113.2, 114.8, 166.2 (C=O), 167.9 (C=O). The *E/Z*-isomers were separated by the procedure of Pfaltz²⁴ using ether–pentane in place of EtOAc–hexane.

Methyl (3S)- or (3R)-3,4-Dimethylpentanoate (33a) and (33b). The procedure of Pfaltz²⁴ was repeated except diglyme was used as the solvent in place of *N,N*-dimethylformamide.

Methyl (2S,3R)- and (2R,3R)-3,4-Dimethyl-2-*n*-hexylpentanoate (34a) and (34b). The procedure described in the preparation of **40e** and **40f** was repeated using 21 mg (0.1 mmol, 1 equiv) of $n\text{-C}_6\text{H}_{13}\text{I}$ and 72 mg (0.5 mmol, 5 equiv) of methyl (3S)-3,4-dimethylpentanoate (**33a**) to afford 20 mg (86%) of methyl (2S,3R)- and (2R,3R)-3,4-dimethyl-2-*n*-hexylpentanoate (**34a**) and (**34b**): IR (CHCl_3) 1730 cm^{-1} ; ^1H NMR (CDCl_3) δ 0.79–0.92 (m, 12, CH_3), 1.20–1.30 (m, 10, CH_2), 1.42–1.75 (m, 2, CH), 2.25–2.40 (m, 1, CHCO_2CH_3), 3.68 (s, 3, OCH_3); exact mass spectrum calcd for $\text{C}_{14}\text{H}_{26}\text{O}$ [$\text{M}^+ - \text{OCH}_3$] 197.1907, found 197.1921; calcd for $\text{C}_{11}\text{H}_{21}\text{O}_2$ [$\text{M}^+ - \text{C}_3\text{H}_7$] 185.1543, found 185.1542. The parent ion was too weak to obtain an exact mass measurement.

Methyl (2R,3S)- and (2S,3S)-3,4-Dimethyl-2-*n*-hexylpentanoate (34c) and (34d). The procedure described in the preparation of **40e** and **40f** was repeated using 21 mg (0.1 mmol, 1 equiv) of $n\text{-C}_6\text{H}_{13}\text{I}$ and 72 mg (0.5 mmol, 5 equiv) of methyl (3R)-3,4-dimethylpentanoate (**33b**) to afford 19 mg (82%) of methyl (2R,3S)- and (2S,3S)-3,4-dimethyl-2-*n*-hexylpentanoate (**34c**) and (**34d**): IR (CHCl_3) 1730 cm^{-1} ; ^1H NMR (CDCl_3) δ 0.79–0.92 (m, 12, CH_3), 1.20–1.30 (m, 10, CH_2), 1.42–1.75 (m, 2, CH), 2.25–2.40 (m, 1, CHCO_2CH_3), 3.68 (s, 3, OCH_3); exact mass spectrum calcd for $\text{C}_{14}\text{H}_{26}\text{O}_2$ [M^+] 228.2089, found 228.2091; calcd for $\text{C}_{11}\text{H}_{25}\text{O}$ [$\text{M}^+ - \text{OCH}_3$] 185.1543, found 185.1544.

Methyl (20R,23S)- and (20R,23R)-4 α -Methyl-27-nor-5 α -cholestane-23-carboxylate (37a) and (37b). The procedure

described for the preparation of **40e** and **40f** was repeated using 58 mg (0.5 mmol, 5 equiv) of methyl pentanoate (**35**) and 44 mg (0.1 mmol, 1 equiv) of **21a** to afford 23 mg (54%) of **37a** and **37b**: IR (CHCl₃) 1730 (C=O) cm⁻¹; ¹H NMR (CDCl₃) δ 0.61 (s, 3, C-18 CH₃), 0.78 (s, 3, C-19 CH₃), 0.81 (d, *J* = 5 Hz, 3, C-4α CH₃), 0.89 (t, *J* = 8.4 Hz, 3, C-26 CH₃), 0.91 (d, *J* = 7.2 Hz, 3, C-21 CH₃), 2.41–2.53 (m, 1, CHCO₂CH₃), 3.66 (s, 3, CO₂CH₃); exact mass spectrum calcd for C₂₉H₅₀O₂ [M⁺] 430.3811, found 430.3823; calcd for ¹³C₂₉H₅₀O₂ [M⁺ + 1] 431.3844, found 431.3843; calcd for C₂₇H₄₈O₂ [M⁺ - C₂H₄] 401.3420, found 401.3420. GC analysis on an SE-30 fused silica gel capillary column (15 m) indicated that the product was a 5.1:1 ratio of **37a** and **37b**.

Methyl (20S,23S)- and (20S,23R)-4α-Methyl-27-nor-5α-cholestane-23-carboxylate (37c) and (37d). The procedure described for the preparation of **40e** and **40f** was repeated using 58 mg (0.5 mmol, 5 equiv) of methyl pentanoate (**35**) and 44 mg (0.1 mmol, 1 equiv) of **21b** to afford 22 mg (51%) of **37c** and **37d**: IR (CHCl₃) 1730 (C=O) cm⁻¹; ¹H NMR (CDCl₃) δ 0.65 (s, 3, C-18 CH₃), 0.78 (s, 3, C-19 CH₃), 0.80 (d, *J* = 4.9 Hz, 3, C-4α CH₃), 0.88–0.92 (m, 6, C-21, C-26 CH₃), 2.33–2.58 (m, 1, CHCO₂CH₃), 3.66 (s, 3, CO₂CH₃); exact mass spectrum calcd for C₂₉H₅₀O₂ [M⁺], found 430.3812. GC analysis on an SE-30 fused silica gel capillary column (15 m) indicated that the product was a 5.3:1 ratio of **37c** and **37d**.

Methyl (20R,23S)- and (20R,23R)-4α-Methyl-5α-cholestane-23-carboxylate (38a) and (38b). The procedure described for the preparation of **40e** and **40f** was repeated using 65 mg (0.5 mmol, 5 equiv) of methyl 4-methylpentanoate (**36**) and 44 mg (0.1 mmol, 1 equiv) of **21a** to afford 25 mg (58%) of **38a** and **38b**: IR (CHCl₃) 1730 (C=O) cm⁻¹; ¹H NMR (CDCl₃) δ 0.62 (s, 3, C-18 CH₃), 0.78 (s, 3, C-19 CH₃), 0.81 (d, *J* = 5 Hz, 3, C-4α CH₃), 0.85 (d, *J* = 6.4 Hz, 6, C-26, C-27 CH₃), 0.91 (d, *J* = 7.2 Hz, 3, C-21 CH₃), 2.40–2.50 (m, 1, CHCO₂CH₃), 3.66 (s, 3, CO₂CH₃); exact mass spectrum calcd for C₃₀H₅₀O₂ [M⁺] 444.3959, found 444.3951. GC analysis on an SE-30 fused silica gel capillary column (15 m) indicated that the product was a 6.6:1 ratio of **38a** and **38b**.

Methyl (20R,23R)- and (20R,23S)-4α-Methyl-24-methyl-ene-5α-cholestane-23-carboxylate (39a) and (39b). The procedure described for the preparation of **40e** and **40f** was repeated using 71 mg (0.5 mmol, 5 equiv) of methyl 3,4-dimethyl-2-pentanoate (**32**; mixture of *E/Z* isomers) and 44 mg (0.1 mmol, 1 equiv) of **21a** to afford 35 mg (76%) of **39a** and **39b** that could not be separated by chromatography: IR (CHCl₃) 1730 (C=O), 890 (C=CH₂) cm⁻¹; ¹H NMR (CDCl₃) δ 0.63 (s, 3, C-18 CH₃), 0.78 (s, 3, C-19 CH₃), 0.79 (d, *J* = 5 Hz, 3, C-4α CH₃), 0.92 (d, *J* = 6.5 Hz, 3, C-21 CH₃), 1.04 (d, *J* = 6.8 Hz, 6, C-26, C-27 CH₃), 3.10–3.21 (m, 1, CHCO₂CH₃), 3.66 (s, 3, CO₂CH₃), 4.88 and 4.94 (two s, 2, C=CH₂); exact mass spectrum calcd for C₃₁H₅₂O₂ [M⁺] 456.3967, found 456.3973; calcd for C₃₀H₄₈O₂ [M⁺ - CH₃] 441.3733, found 441.3743. GC analysis on an SE-30 fused silica gel capillary column (15 m) indicated that the product was a 2.4:1 ratio of **39a** and **39b**.

Methyl (20S,23R)- and (20S,23S)-4α-Methyl-24-methyl-ene-5α-cholestane-23-carboxylate (39c) and (39d). The procedure described for the preparation of **40e** and **40f** was repeated using 71 mg (0.5 mmol, 5 equiv) of methyl 3,4-dimethyl-2-pentanoate (**32**; mixture of *E/Z* isomers) and 44 mg (0.1 mmol, 1 equiv) of **21b** to afford 33 mg (72%) of **39c** and **39d**: IR (CHCl₃) 1730 (C=O), 890 cm⁻¹; ¹H NMR (CDCl₃) δ 0.65 (s, 3, C-18 CH₃), 0.78 (s, 3, C-19 CH₃), 0.79 (d, *J* = 5 Hz, 3, C-4α CH₃), 0.92 (d, *J* = 6.4 Hz, 3, C-21 CH₃), 1.04 (d, *J* = 6.8 Hz, 6, C-26, C-27 CH₃), 3.08–3.18 (m, 1, CHCO₂CH₃), 3.65 (s, 3, CO₂CH₃), 4.93 and 4.99 (two s, 2, C=CH₂); exact mass spectrum calcd for C₃₁H₅₂O₂ [M⁺] 456.3967, found 456.3978. GC analysis on an SE-30 fused silica gel capillary column (15 m) indicated that the product was a 2.2:1 ratio of **39c** and **39d**.

Methyl (20R,23R,24S)- and (20R,23S,24S)-4α,24-Dimethyl-5α-cholestane-23-carboxylate (40a) and (40b). The procedure described for the preparation of **40e** and **40f** was repeated using 44 mg (0.1 mmol) of **21a** with 72 mg (0.5 mmol) of **33b** to afford 38 mg (83%) of a mixture of **40a** and **40b** as an oil: IR (CHCl₃) 1730 (C=O), 1460, 1430, 1360, 1165 cm⁻¹; ¹H NMR (CDCl₃) δ 0.64 (s, 3, C-18 CH₃), 0.72–0.92 (m, 18, C-4α, C-19 and side-chain CH₃), 3.65 and 3.77 (two s, 3, OCH₃); exact mass spectrum calcd for C₃₀H₅₄O₂ 458.4124, found 458.4127. GC analysis on an SE-30

fused silica gel capillary column (15 m) indicated that the product was a 4:1 ratio of **40a** and **40b**.

Methyl (20R,23R,24R)- and (20R,23S,24R)-4α,24-Dimethyl-5α-cholestane-23-carboxylate (40e) and (40f). To a solution of 105 μL (76 mg, 0.75 mmol, 7.5 equiv) of diisopropylamine in 0.5 mL of anhydrous THF at -20 °C was added 200 μL (0.5 mmol, 5 equiv) of 2.5 M *n*-BuLi in hexane. The solution was stirred at -20 °C for 20 min and cooled to -78 °C. To this solution was added dropwise 72 mg (0.5 mmol, 5 equiv) of methyl (3S)-3,4-dimethylpentanoate (**33a**) in 0.5 mL of THF. The mixture was stirred at -78 °C for 30 min, and 44 mg (0.1 mmol, 1 equiv) of **21a** in 0.5 mL of THF and 87 μL (0.5 mmol, 5 equiv) of anhydrous ((CH₃)₂N)₃PO was added dropwise. The mixture was stirred at -78 °C for 4 h and was allowed to warm to 25 °C. The mixture was diluted with hexane and water and extracted with hexane. The combined organic extracts were washed successively with saturated NH₄Cl solution and brine and dried over anhydrous MgSO₄. The crude product was chromatographed on silica gel using 1:50 EtOAc-hexane to afford 37 mg (81%) of **40e** and **40f** as an oil: IR (CHCl₃) 1730 (C=O), 1460, 1435, 1370, 1160 cm⁻¹; ¹H NMR (CDCl₃) δ 0.63 (s, 3, C-18 CH₃), 0.75–0.93 (m, 18, C-4α, C-19, and side-chain CH₃), 3.64 and 3.65 (two s, 3, OCH₃); exact mass spectrum calcd for C₃₀H₅₄O₂ 458.4124, found 458.4115; calcd for [M⁺ - CH₃] C₃₀H₅₁O₂ 443.3889, found 443.3881. GC analysis on an SE-30 fused silica gel capillary column (15 m) indicated that the product was a 1:4 ratio of **40e** and **40f**.

(20R,23R,24S)- and (20R,23S,24S)-5α-Dinosteran-29-ol (41a) and (41b). The reduction of a mixture of **40a** and **40b** with LiAlH₄ provided a mixture of **41a** and **41b**, respectively, that was separated by chromatography on silica gel using 1:10 EtOAc-hexane. The principal epimer **41a** had the following physical and spectral data: mp 162–164 °C; IR (CHCl₃) 3615, 1460, 1440, 1380 cm⁻¹; ¹H NMR (CDCl₃) δ 0.65 (s, 3, C-18 CH₃), 0.75 (d, *J* = 6.9 Hz, 3, C-28 CH₃), 0.78 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 0.88 (d, *J* = 6.6 Hz, 3, C-21 CH₃), 0.92 (d, *J* = 6.4 Hz, 6, C-26 and C-27 CH₃), 3.56 (ddd, *J*₁ = 3.6 Hz, *J*₂ = 7.9 Hz, *J*₃ = 10 Hz, 2, CHCH₂OH); ¹³C NMR (CDCl₃) δ 11.4, 12.1, 13.3, 18.8, 19.9, 20.6, 21.0, 21.6, 21.9, 24.1, 24.3, 28.8, 29.8, 31.3, 32.2, 32.7, 34.1, 35.1, 36.4, 36.6, 38.9 (two C), 39.0, 40.2, 42.6, 53.8, 54.8, 56.8, 57.3, 63.9 (CH₂OH); exact mass spectrum calcd for C₃₀H₅₄O [M⁺] 430.4175, found 430.4171. The minor epimer **41b** had the following physical and spectral data: mp 177–179 °C; IR (CHCl₃) 3615, 1460, 1440, 1380 cm⁻¹; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.75 (d, *J* = 7 Hz, 3, C-28 CH₃), 0.79 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 0.79 (s, 3, C-19 CH₃), 0.89 (d, *J* = 6.6 Hz, 3, C-21 CH₃), 0.93 (d, *J* = 6.9 Hz, 6, C-26 and C-27 CH₃), 3.50 (ddd, *J*₁ = 4 Hz, *J*₂ = 8.5 Hz, *J*₃ = 10 Hz, 2, CH₂OH); ¹³C NMR (CDCl₃) δ 11.8, 12.2, 13.3, 18.5, 20.5, 21.0, 21.1, 21.5, 21.9, 24.2, 24.3, 28.6, 31.1, 31.4, 32.3, 33.5, 35.2, 36.4, 36.5, 36.6, 38.8, 38.9, 39.5, 40.3, 42.7, 53.6, 54.9, 56.8, 57.4, 64.2 (CH₂OH); exact mass spectrum calcd for C₃₀H₅₄O [M⁺] 430.4175, found 430.4173.

(20R,23R,24R)- and (20R,23S,24R)-5α-Dinosteran-29-ol (41e) and (41f). The reduction of a mixture of **40e** and **40f** with LiAlH₄ provided a mixture of **41e** and **41f**, respectively, that was separated by chromatography on silica gel using 1:10 EtOAc-hexane. The principal epimer **41f** had the following physical and spectral data: mp 158–162 °C; IR (CHCl₃) 3615, 1460, 1440, 1380 cm⁻¹; ¹H NMR (CDCl₃) δ 0.65 (s, 3, C-18 CH₃), 0.79 (two d, *J* = 6.6 Hz, 6, C-4α, C-28 CH₃), 0.79 (s, 3, C-19 CH₃), 0.85 (d, *J* = 6.6 Hz, 3, C-21 CH₃), 0.92 (d, *J* = 6.6 Hz, 6, C-26, C-27 CH₃), 3.52 (d, *J* = 6.2 Hz, 2, CH₂OH); ¹³C NMR (CDCl₃) δ 12.0 (two C), 13.3, 19.5, 19.7, 20.6, 21.0, 21.9, 22.3, 24.2, 24.3, 28.5, 29.4, 31.3, 32.2, 34.4, 35.1, 36.0, 36.4, 36.5, 38.8, 39.4, 40.2, 41.4, 42.5, 53.5, 54.7, 56.6, 57.8, 65.6 (CH₂OH); exact mass spectrum calcd for C₃₀H₅₄O [M⁺] 430.4175, found 430.4175. The minor epimer **41e** had the following physical and spectral data: mp 128–130 °C; IR (CHCl₃) 3615 (OH), 1460, 1440, 1380 cm⁻¹; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.78 (d, *J* = 6.6 Hz, 3, C-4α CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6.3 Hz, 3, C-28 CH₃), 0.82 (d, *J* = 6.9 Hz, 3, C-21 CH₃), 0.91 (d, *J* = 6.4 Hz, 3, C-26 CH₃), 0.93 (d, *J* = 6.6 Hz, 3, C-27 CH₃), 3.64 (d, *J* = 2.8 Hz, 2, CH₂OH); ¹³C NMR (CDCl₃) δ 11.7, 12.1, 13.2, 17.6, 18.8, 20.6, 21.0, 21.9, 22.1, 24.2, 24.3, 28.8, 28.9, 31.3, 32.2, 34.7, 35.1, 36.4, 36.5, 38.8, 39.9, 40.2, 40.9, 42.6, 53.5, 54.8, 56.8, 57.3, 62.8 (CH₂OH); exact mass spectrum calcd for C₃₀H₅₄O [M⁺] 430.4175, found 430.4177; calcd for ¹³C₂₉H₅₄O [M⁺ + 1] 431.4208, found 431.4219; calcd for

$C_{29}H_{51}O$ [$M^+ - CH_3$] 415.3940, found 415.3949; calcd for $C_{28}H_{49}O$ [$M^+ - C_2H_5$] 401.3783, found 401.3782.

(20R,23R,24S)-5 α -Dinosteran-29-ol Mesylate. The procedure of Crossland¹⁸ was repeated using 118 mg (0.27 mmol) of **41a** and 65 mg (0.54 mmol, 2 equiv) of CH_3SO_2Cl to afford 129 mg (93%) of mesylate: mp 125.5–126.5 °C; IR (CHCl₃) 1460, 1440, 1360, 1340, 1170, 940, 830 cm^{-1} ; ¹H NMR (CDCl₃) δ 0.65 (s, 3, C-18 CH₃), 0.76 (d, $J = 6.8$ Hz, C-28 CH₃), 0.77 (d, $J = 6.3$ Hz, C-4 α CH₃), 0.78 (s, 3, C-19 CH₃), 0.87 (d, $J = 6.6$ Hz, 3, C-21 CH₃), 0.90 (d, $J = 6.6$ Hz, 3, C-26 CH₃), 0.92 (d, $J = 6.3$ Hz, 3, C-27 CH₃), 2.99 (s, 3, OSO₂CH₃), 4.12 (ddd, $J_1 = 4.1$ Hz, $J_2 = 7.9$ Hz, $J_3 = 9.7$ Hz, 2, CH₂OSO₂CH₃); ¹³C NMR (CDCl₃) δ 11.3, 12.1, 13.2, 18.4, 19.8, 20.5, 20.9, 21.4, 21.8, 24.0, 24.2, 28.6, 29.6, 31.2, 32.2, 32.3, 33.8, 35.0, 36.0, 36.4, 36.5, 37.2, 38.8, 39.0, 40.1, 42.5, 53.4, 54.6, 56.7, 57.2, 70.9 (CH₂OSO₂CH₃).

(20R,23S,24R)-5 α -Dinosteran-29-ol Mesylate. The procedure of Crossland¹⁸ was repeated using 23 mg (0.05 mmol) of **41f** and 12 mg (0.1 mmol, 2 equiv) of CH_3SO_2Cl to afford 25 mg (93%) of mesylate: mp 154–155 °C; IR (CHCl₃) 1460, 1440, 1360, 1340, 1170, 940, 830 cm^{-1} ; ¹H NMR (CHCl₃) δ 0.64 (s, 3, C-18 CH₃), 0.79 (d, $J = 6.6$ Hz, 3, C-4 α CH₃), 0.79 (s, 3, C-19 CH₃), 0.81 (d, $J = 6.6$ Hz, 3, C-28 CH₃), 0.85 (d, $J = 6.6$ Hz, 3, C-21 CH₃), 0.92 (d, $J = 6.6$ Hz, 3, C-26 CH₃), 0.94 (d, $J = 6.4$ Hz, 3, C-27 CH₃), 3.00 (s, 3, SO₂CH₃), 4.10 (d, $J = 6.5$ Hz, 2, CH₂OSO₂CH₃); ¹³C NMR (CDCl₃) δ 11.9, 12.0, 13.2, 19.3, 19.5, 20.6, 20.9, 21.9, 22.1, 24.1, 24.2, 28.5, 29.2, 31.3, 32.2, 34.0, 35.1, 36.0, 36.4, 36.5, 37.3, 38.5, 38.8, 39.4, 40.1, 42.6, 53.4, 54.7, 56.6, 57.6, 72.4 (CH₂OSO₂CH₃).

(20R,23S,24R)-5 α -Dinosterane (2a). To 18 mg (0.47 mmol) of LiAlH₄ in 0.5 mL of anhydrous THF under N₂ was added 25 mg (0.049 mmol) of (20R,23S,24R)-5 α -dinosteran-29-ol mesylate in 0.5 mL of THF. The mixture was refluxed for 5 h. The solution was cooled to 0 °C, quenched with water, and diluted with hexane. The organic solution was washed with brine and dried over anhydrous MgSO₄. The product was chromatographed on silica gel using hexane to afford 20 mg (96%) of **2a**: mp 106.5–107 °C; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.69 (d, $J = 7.0$ Hz, 3, C-28 CH₃), 0.69 (d, $J = 6.5$ Hz, 3, C-29 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, $J = 6.5$ Hz, 3, C-4 α CH₃), 0.87 (d, $J = 6.5$ Hz, 3, C-21 CH₃), 0.88 (d, $J = 6.5$ Hz, 3, C-27 CH₃), 0.89 (d, $J = 6.4$ Hz, 3, C-26 CH₃); ¹³C NMR (CDCl₃) δ 12.0, 12.1, 13.3, 18.8, 20.1 (two C), 20.6, 21.0, 21.9, 22.6, 24.2, 24.3, 28.5, 28.6, 31.2, 32.2, 34.6, 35.1, 35.9, 36.4, 36.6, 38.8, 40.2, 40.6, 42.5, 44.0, 53.5, 54.8, 56.6, 57.9.

(20R,23R,24S)-5 α -Dinosterane (2d). The procedure described for the preparation of **2a** was repeated using 90 mg (2.38 mmol) of LiAlH₄ in 1.3 mL of anhydrous THF and 129 mg (0.25 mmol) of the (20R,23R,24S)-5 α -dinosteran-29-ol mesylate to afford, after chromatography on silica gel using hexane, 101 mg (96%) of **2d**: mp 129.5–130 °C; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.70 (d, $J = 6.8$ Hz, 3, C-29 CH₃), 0.72 (d, $J = 6.8$ Hz, 3, C-28 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, $J = 6.4$ Hz, 3, C-4 α CH₃), 0.81 (d, $J = 6.8$ Hz, 3, C-21 CH₃), 0.87 (d, $J = 6.4$ Hz, 3, C-27 CH₃), 0.88 (d, $J = 6.8$ Hz, 3, C-26 CH₃); ¹³C NMR (CDCl₃) δ 11.4, 12.1, 13.3, 18.349, 18.381, 19.1, 20.6, 21.0, 21.8, 21.9, 24.1, 24.3, 28.6, 29.5, 31.28, 31.29, 32.2, 33.7, 35.1, 36.4, 36.6, 38.1, 38.8, 40.2, 42.6, 45.2, 53.5, 54.8, 56.8, 57.4.

(20R,23R)- and (20R,23S)-5 α -Dinoster-24(28)-en-29-ol (42a) and (42b). To a solution of 32 mg (0.07 mmol) of a mixture of **39a** and **39b** in 3 mL of anhydrous THF was added 8 mg (0.21 mmol, 3 equiv) of LiAlH₄. The solution was stirred for 2 h and quenched with H₂O. The product was extracted with Et₂O, dried over anhydrous MgSO₄, and chromatographed on silica gel using 1:10 EtOAc–hexane to afford 21 mg (67%) of **42a** and 9 mg (29%) of **42b**. The epimer **42a** had the following spectral data: IR (CHCl₃) 3430 (OH), 890 cm^{-1} ; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.79 (d, $J = 6.3$ Hz, 3, C-4 α CH₃), 0.79 (s, 3, C-19 CH₃), 0.92 (d, $J = 7.7$ Hz, 3, C-21 CH₃), 1.06 (d, $J = 7$ Hz, 6, C-26, C-27 CH₃), 3.47–3.66 (m, 2, CH₂OH), 4.80 and 4.96 (two s, 2, C=CH₂); ¹³C NMR (CDCl₃) δ 12.1 (C-18), 13.2 (C-19), 18.9, 20.6, 20.9, 21.9, 22.4, 22.5, 24.1, 24.2, 28.6, 31.3, 32.2, 33.7, 34.4, 35.1, 36.4, 36.5, 38.7, 38.8, 40.2, 42.6, 43.8, 53.5, 54.7, 56.7, 57.0, 63.7 (CH₂OH), 107.2 (C=CH₂), 158.4 (C=CH₂); exact mass spectrum calcd for $C_{30}H_{52}O$ [M^+] 428.4018, found 428.4019. The epimer **42b** had the following spectral data: IR (CHCl₃) 3430 (OH), 890 cm^{-1} ; ¹H NMR (CDCl₃) δ 0.61 (s, 3, C-18 CH₃), 0.78 (s, 3, C-19 CH₃), 0.79

(d, $J = 6.3$ Hz, 3, C-4 α CH₃), 0.91 (d, $J = 6.3$ Hz, 3, C-21 CH₃), 1.04 (d, $J = 6.2$ Hz, 6, C-26, C-27 CH₃), 3.39–3.55 (m, 2, CH₂OH), 4.82 and 5.01 (two s, 2, C=CH₂); ¹³C NMR (CDCl₃) δ 11.1 (C-18), 12.4 (C-19), 17.9, 19.7, 20.1, 21.0, 21.6, 22.2, 23.3, 23.4, 27.6, 30.5, 31.4, 32.1, 32.6, 34.3, 35.6, 35.8, 36.7, 38.1, 39.4, 41.8, 44.5, 52.7, 54.0, 56.0, 66.8 (CH₂OH), 108.4 (C=CH₂), 156.1 (C=CH₂); exact mass spectrum calcd for $C_{30}H_{52}O$ [M^+] 428.4018, found 428.4013.

(20R,23S)-5 α -Dinoster-24(28)-en-29-ol *tert*-Butyldimethylsilyl Ether. The procedure of Corey³⁰ was repeated using 38 mg (0.089 mmol) of **42b**, 59 mg (0.89 mmol, 10 equiv) of imidazole, and 68 mg (0.45 mmol, 5 equiv) of *tert*-butyldimethylsilyl chloride in 4 mL of *N,N*-dimethylformamide for 30 min at 60 °C to afford, after chromatography on silica gel using hexane, 47 mg (96%) of (20R,23S)-5 α -dinoster-24(28)-en-29-ol *tert*-butyldimethylsilyl ether: mp 144–145 °C; IR (CHCl₃) 1460, 1440, 1380, 1250, 1050, 840 cm^{-1} ; ¹H NMR (CDCl₃) δ 0.03 (s, 6, Si(CH₃)₂), 0.63 (s, 3, C-18 CH₃), 0.79 (d, $J = 6.4$ Hz, 3, C-4 α CH₃), 0.79 (s, 3, C-19 CH₃), 0.88 (d, $J = 8.1$ Hz, 3, C-21 CH₃), 0.89 (s, 9, C(CH₃)₃), 1.01 (d, $J = 6.5$ Hz, 3, C-26 CH₃), 1.04 (d, $J = 6.6$ Hz, 3, C-27 CH₃), 3.37 (ddd, $J_1 = 5.3$ Hz, $J_2 = 8$ Hz, $J_3 = 10$ Hz, 2, CH₂OTBDMS), 4.68 (s, 1, vinylic H), 4.84 (s, 1, vinylic H); ¹³C NMR (CDCl₃) δ -5.5 (Si(CH₃)₂), 12.0, 13.3, 18.3, 18.8, 20.6, 21.0, 21.9, 22.1, 22.5, 24.1, 24.3, 26.0 (C(CH₃)₃), 28.4, 31.3, 32.3, 33.2, 34.3, 35.1, 36.4, 36.6, 37.8, 38.9, 40.2, 42.5, 44.6, 53.5, 54.8, 56.8, 57.2, 68.4 (CH₂-OTBDMS), 107.2 (C=CH₂), 157.1 (C=CH₂); exact mass spectrum calcd for $C_{32}H_{57}OSi$ 485.4179, found 485.4178.

(20R,23R)-5 α -Dinoster-24(28)-en-29-ol *tert*-Butyldimethylsilyl Ether. The procedure of Corey³⁰ was repeated using 78 mg (0.18 mmol) of **42a**, 119 mg (1.8 mmol, 10 equiv) of imidazole, and 136 mg (0.9 mmol, 5 equiv) of *tert*-butyldimethylsilyl chloride in 8 mL of *N,N*-dimethylformamide for 30 min at 60 °C to afford, after chromatography on silica gel using hexane, 96 mg (96%) of (20R,23R)-5 α -dinoster-24(28)-en-29-ol *tert*-butyldimethylsilyl ether: mp 130.5–131 °C; IR (CHCl₃) 1460, 1440, 1380, 1250, 1090, 840 cm^{-1} ; ¹H NMR (CDCl₃) δ 0.03 (s, 6, Si(CH₃)₂), 0.65 (s, 3, C-18 CH₃), 0.78 (s, 3, C-19 CH₃), 0.79 (d, $J = 6.3$ Hz, 3, C-4 α CH₃), 0.87 (s, 9, C(CH₃)₃), 0.92 (d, $J = 6.4$ Hz, 3, C-21 CH₃), 1.02 (d, $J = 6.7$ Hz, 3, C-26 and C-27 CH₃), 3.52 (d, $J = 5.8$ Hz, 2, CH₂OTBDMS), 4.70 (s, 1, vinylic H), 4.78 (s, 1, vinylic H); ¹³C NMR (CDCl₃) δ -5.4 (Si(CH₃)₂), 12.1, 13.3, 18.2, 19.2, 20.6, 21.0, 21.9, 22.1, 22.2, 24.2, 24.3, 25.9 (C(CH₃)₃), 28.5, 31.3, 32.3, 34.7, 34.72, 35.1, 36.4, 36.6, 38.9, 40.0, 40.2, 42.6, 43.5, 53.5, 54.8, 56.7, 57.4, 66.8, (CH₂-OTBDMS), 106.0 (C=CH₂), 159.5 (C=CH₂); exact mass spectrum calcd for $C_{32}H_{57}OSi$ 485.4179, found 485.4176.

(20R,23R,24R)- and (20R,23R,24S)-5 α -Dinosteran-29-ol *tert*-Butyldimethylsilyl Ether. A mixture of 96 mg (0.22 mmol) of (20R,23R)-5 α -dinoster-24(28)-en-29-ol *tert*-butyldimethylsilyl ether (prepared from **42a**) and 45 mg (0.2 mmol) of PtO₂ in 3 mL of hexane was hydrogenated at 60 psi for 3 h at 25 °C. The catalyst was removed by filtration, and the product was concentrated to afford 94 mg (98%) of a mixture of saturated TBDMS ethers, (20R,23R,24R)- and (20R,23R,24S)-5 α -dinosteran-29-ol *tert*-butyldimethylsilyl ether. The mixture was chromatographed on silica gel using hexane to afford (20R,23R,24R)-5 α -dinosteran-29-ol *tert*-butyldimethylsilyl ether as the principal isomer: mp 142–142.5 °C; IR (CHCl₃) 1465, 1375, 1250, 1090, 840 cm^{-1} ; ¹H NMR (CDCl₃) δ 0.02 (s, 6, Si(CH₃)₂), 0.65 (s, 3, C-18 CH₃), 0.76 (d, $J = 6.6$ Hz, 3, C-28 CH₃), 0.77 (d, $J = 6.6$ Hz, 3, C-4 α CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, $J = 8.1$ Hz, 3, C-21 CH₃), 0.88 (s, 9, SiC(CH₃)₃), 0.88 (d, $J = 6.4$ Hz, 3, C-26 CH₃), 0.89 (d, $J = 6.6$ Hz, 3, C-27 CH₃), 3.70 (ddd, $J_1 = 4$ Hz, $J_2 = 3.8$ Hz, $J_3 = 10$ Hz, 2, CH₂OTBDMS); ¹³C NMR (CDCl₃) δ -5.54 (Si(CH₃)₂), -5.49 (Si(CH₃)₃), 11.7, 12.2, 13.3, 18.0, 18.2, 18.8, 20.6, 21.0, 21.9, 22.2, 24.2, 24.3, 25.9 (C(CH₃)₃), 28.5, 29.0, 31.3, 32.3, 34.3, 35.1, 36.5, 36.6, 36.9, 38.9, 39.6, 40.3, 41.3, 42.6, 53.5, 54.8, 56.8, 57.5, 62.4 (CH₂OTBDMS). The minor isomer, (20R,23R,24S)-5 α -dinosteran-29-ol *tert*-butyldimethylsilyl ether, had the following physical and spectral data: mp 144.5–145 °C; IR (CHCl₃) 1460, 1380, 1250, 1090, 840 cm^{-1} ; ¹H NMR (CDCl₃) δ 0.03 (s, 6, Si(CH₃)₂), 0.65 (s, 3, C-18 CH₃), 0.70 (d, $J = 6.7$ Hz, 3, C-28 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, $J = 6.3$ Hz, 3, C-4 α CH₃), 0.84 (d, $J = 6.6$ Hz, 3, C-21 CH₃), 0.86–0.92 (m, 6, C-26 and C-27 CH₃), 0.88 (s, 9, SiC(CH₃)₃), 3.47 (ddd, $J_1 = 4$ Hz, $J_2 = 7$ Hz, $J_3 = 10$)

H_z, 2, CH₂OTBDMS); ¹³C NMR (CDCl₃) δ -5.6 (SiCH₃), 11.3, 12.0, 13.1, 18.1, 18.7, 19.6, 20.5, 20.9, 21.6, 21.8, 24.0, 24.2, 25.8 (C(CH₃)₃), 28.6, 29.6, 31.2, 32.2, 33.0, 33.9, 35.0, 36.4, 36.5, 38.8 (two C), 39.0, 40.2, 42.5, 53.5, 54.8, 57.5, 64.0 (CH₂OTBDMS); exact mass spectrum calcd for C₃₂H₅₀OSi 487.4335, found 487.4337.

(20R,23S,24S)- and (20R,23S,24R)-5α-Dinosteran-29-ol tert-Butyldimethylsilyl Ether. A mixture of 47 mg (0.086 mmol) of (20R,23S)-5α-dinoster-24(28)-en-29-ol tert-butyldimethylsilyl ether (prepared from 42b) and 23 mg (0.1 mmol) of PtO₂ was hydrogenated in 2 mL of hexane at 60 psi for 3 h at 25 °C. The catalyst was removed by filtration, and the product was concentrated to afford 47 mg (100%) of a mixture of saturated TBDMS ethers, (20R,23S,24S)- and (20R,23S,24R)-5α-dinosteran-29-ol tert-butyldimethylsilyl ether. This mixture was not separable by chromatography on silica gel and was used directly in the deprotection step described below.

(20R,23R,24R)-5α-Dinosteran-29-ol (41e). To a solution of 69 mg (0.13 mmol) of (20R,23R,24R)-5α-dinosteran-29-ol tert-butyldimethylsilyl ether in 5 mL of THF was added 278 μL (0.38 mmol, 3 equiv) of 1 M (n-Bu)₄NF in THF. The solution was allowed to stir for 24 h at 25 °C and was diluted with EtOAc. The solution was washed with water and brine and dried over anhydrous MgSO₄. The product was chromatographed on silica gel using 1:10 EtOAc-hexane to afford 51 mg (95%) of 41e having spectral data in agreement with that reported in the preparation of 41e from 40e (vide supra).

(20R,23S,24S)-5α-Dinosteran-29-ol (41b). The procedure described for the preparation of 41e was repeated using 47 mg (0.09 mmol) of an inseparable mixture of (20R,23S,24S)- and (20R,23S,24R)-5α-dinosteran-29-ol tert-butyldimethylsilyl ethers and 257 μL (0.26 mmol, 3 equiv) of 1 M (n-Bu)₄NF in 4 mL of THF to afford, after chromatography on silica gel using 1:10 EtOAc-hexane, 35 mg (94%) of 41b and 41f, respectively. This mixture was separated by HPLC on Beckman C₁₈ reversed-phase column using 2:10:100 water-EtOAc-MeOH to give a pure sample of 41b having spectral data in agreement with that reported in the preparation of 41b from 40b (vide supra).

(20R,23R,24R)-5α-Dinosteran-29-ol Mesylate. The procedure of Crossland¹⁸ was repeated using 51 mg (0.12 mmol) of 41e and 27 mg (0.23 mmol, 2 equiv) of CH₃SO₂Cl to afford 59 mg (98%) of (20R,23R,24R)-5α-dinosteran-29-ol mesylate: mp 116–116.5 °C; IR (CHCl₃) 1460, 1440, 1360, 1170, 970, 940, 830 cm⁻¹; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6.5 Hz, 6, C-4α and C-28 CH₃), 0.83 (d, *J* = 8.1 Hz, 3, C-21 CH₃), 0.91 (d, *J* = 6.3 Hz, 3, C-26 CH₃), 0.93 (d, *J* = 6.7 Hz, 3, C-27 CH₃), 2.99 (s, 3, CH₂OSO₂CH₃), 4.20 (ddd, *J*₁ = 4.5 Hz, *J*₂ = 4 Hz, *J*₃ = 10 Hz, 2, CH₂OSO₂CH₃); ¹³C NMR (CDCl₃) δ 11.6, 12.1, 13.2, 17.8, 18.6, 20.6, 20.9, 21.9 (two C), 24.1, 24.2, 28.5, 29.0, 31.2, 32.2, 34.2, 35.0, 36.4, 36.5 (two C), 37.17, 37.24, 38.8, 40.2, 41.1, 42.6, 53.4, 54.7, 56.7, 57.2, 70.1 (CH₂OSO₂CH₃).

(20R,23S,24S)-5α-Dinosteran-29-ol Mesylate. The procedure of Crossland¹⁸ was repeated using 25 mg (0.06 mmol) of 41b and 13 mg (0.11 mmol, 2 equiv) of CH₃SO₂Cl to afford 29 mg (99%) of (20R,23S,24S)-5α-dinosteran-29-ol mesylate: mp 151–152 °C; IR (CHCl₃) 1460, 1440, 1360, 1340, 1170, 940, 830 cm⁻¹; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6.6 Hz, 6, C-4α CH₃ and C-28 CH₃), 0.84–0.95 (m, 9, C-21, C-26 and C-27 CH₃), 3.00 (s, 3, CH₂OSO₂CH₃), 4.08 (septet, *J*₁ = 2 Hz, *J*₂ = 9 Hz, *J*₃ = 11 Hz, 2, CH₂OSO₂CH₃); ¹³C NMR

(CDCl₃) δ 11.7, 12.1, 13.3, 18.2, 20.6, 21.0, 21.3, 21.9, 24.1, 24.2, 28.7, 31.0, 31.3, 33.3, 35.0, 35.6, 35.9, 36.4, 36.5, 37.2, 38.8, 39.0, 40.2, 42.6, 53.4, 54.7, 56.7, 57.0, 71.7 (CH₂OSO₂CH₃).

(20R,23R,24R)-5α-Dinosterane (2b). The procedure described for the preparation of 2a was repeated using 35 mg (0.92 mmol) of LiAlH₄ in 0.5 mL of anhydrous THF and 50 mg (0.098 mmol) of (20R,23R,24R)-5α-dinosteran-29-ol mesylate to afford, after chromatography on silica gel using hexane, 38 mg (94%) of 2b: mp 143.5–144 °C; ¹H NMR (CDCl₃) δ 0.65 (s, 3, C-18 CH₃), 0.76 (d, *J* = 6.8 Hz, 3, C-28 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6.4 Hz, 3, C-4α CH₃), 0.79 (d, *J* = 6.7 Hz, 3, C-21 CH₃), 0.87 (d, *J* = 6.7 Hz, 3, C-27 CH₃), 0.89 (d, *J* = 6.8 Hz, 3, C-26 CH₃), 0.92 (d, *J* = 6.8 Hz, 3, C-29 CH₃); ¹³C NMR (CDCl₃) δ 11.4, 12.1, 13.3, 14.2, 18.7, 19.4, 20.6, 21.0, 21.7, 21.9, 24.2, 24.3, 28.6, 30.1, 31.2, 31.3, 32.3, 34.1, 35.1, 36.4, 36.6, 38.9, 40.2, 42.6, 42.8, 45.3, 53.5, 54.8, 56.8, 57.3.

(20R,23S,24S)-5α-Dinosterane (2c). The procedure described for the preparation of 2a was repeated using 18 mg (0.47 mmol) of LiAlH₄ in 0.5 mL of anhydrous THF and 25 mg (0.049 mmol) of (20R,23S,24S)-5α-dinosteran-29-ol mesylate to afford, after chromatography on silica gel using hexane, 19 mg (93%) of 2c: mp 161.5–162.5 °C; ¹H NMR (CDCl₃) δ 0.66 (s, 3, C-18 CH₃), 0.72 (d, *J* = 6.8 Hz, 3, C-28 CH₃), 0.79 (s, 3, C-19 CH₃), 0.79 (d, *J* = 6.3 Hz, 3, C-4α CH₃), 0.81 (d, *J* = 6.8 Hz, 3, C-29 CH₃), 0.81 (d, *J* = 6.8 Hz, 3, C-27 CH₃), 0.87 (d, *J* = 6.4 Hz, 3, C-21 CH₃), 0.88 (d, *J* = 6.8 Hz, 3, C-26 CH₃); ¹³C NMR (CDCl₃) δ 10.6, 12.1, 13.3, 15.2, 18.5, 20.6, 21.0, 21.2, 21.5, 21.9, 24.2, 24.3, 28.5, 29.6, 31.0, 31.3, 32.2, 33.3, 35.1, 36.4, 36.6, 38.8, 40.1, 40.2, 42.0, 42.6, 53.5, 54.8, 56.7, 57.4.

Acknowledgment. We acknowledge support of the University of Kentucky Major Research Instrumentation Bond Program in the purchase of equipment used in this study (bond ID no. 7E-8E48-25). One of us (D.S.W.) thanks Chevron Petroleum Technology Co. for their generous financial support. We thank Dr. Nacer-Eddine Slougui for preliminary experiments on the lithocholic acid route, Professor Carl Djerassi for a sample of dinosterol, Dr. Mark Sabol for work on the conversion of dinosterol to 5α-dinosterane, Mr. Claude Dungan for determining NMR spectra, and Professor Andreas Pfaltz for providing the semicorrin catalyst.

Supplementary Material Available: ¹H and ¹³C NMR spectra for (20S)-20-(hydroxymethyl)-4α-methyl-5α-pregnane (20a), (20R)-20-(hydroxymethyl)-4α-methyl-5α-pregnane (20b), mesylate of (20R,23R,24S)-5α-dinosteran-29-ol (41a), mesylate of (20R,23S,24S)-5α-dinosteran-29-ol (41b), mesylate of (20R,23R,24R)-5α-dinosteran-29-ol (41e), mesylate of (20R,23S,24R)-5α-dinosteran-29-ol (41f), tert-butyldimethylsilyl ether of (20R,23S,24S)-5α-dinosteran-29-ol (41b) (¹H NMR only), tert-butyldimethylsilyl ether of (20R,23R,24R)-5α-dinosteran-29-ol (41e), (20R,23S,24R)-5α-dinosterane (2a), (20R,23R,24R)-5α-dinosterane (2b), (20R,23S,24S)-5α-dinosterane (2c), and (20R,23R,24S)-5α-dinosterane (2d) (23 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.